

## PREVALENCE AND RISK FACTORS OF SUBCLINICAL MILK FEVER AND KETOSIS IN LACTATING CROSS-BRED DAIRY COWS WITH THEIR THERAPEUTIC MANAGEMENT IN BANGLADESH

L. Naher,<sup>1\*</sup> M. A. Samad,<sup>1\*\*</sup> S. H. M. F. Siddiki<sup>2</sup> and M. T. Islam<sup>1</sup>

<sup>1</sup>Department of Medicine, Faculty of Veterinary Science, Bangladesh Agricultural University, Mymensingh-2202, Bangladesh. <sup>2</sup>Department of Medicine, Faculty of Veterinary Medicine and Animal Science, BSMRAU, Gazipur-1706, Bangladesh \*Part of MS thesis \*\*E-mail: [vetmedbd@yahoo.com](mailto:vetmedbd@yahoo.com)

### ABSTRACT

**Background:** Bovine Milk fever (MF/hypocalcaemia) and ketosis (CK/hypoglycemia and hyperketonemia) both in clinical and sub-clinical forms are the most important metabolic diseases caused by metabolic disorders of calcium and carbohydrate respectively that affect mainly high milk yielding dairy cows worldwide. Sub-clinical form may be more costly due to comparatively high prevalence and consequence of high risk of decreased productive and reproductive performances with increased reproductive and other disorders.

**Objectives:** The objectives of this study were to determine the prevalence of sub-clinical hypocalcaemia (SCHC) and sub-clinical ketosis (SCK) and to investigate important potential risk factors for SCHC and SCK with their therapeutic management in lactating cross-bred dairy cows.

**Materials and Methods:** A cross sectional study was conducted on 220 dairy crossbred (HF × L = 190, SH × L = 20 and JS × L = 10) cows maintained in nine dairy farms and one smallholder farm during the period from July to November 2016. The parity (1 to 8), lactation stages (1 to 13 weeks), body condition score (BCS), breed (3 crossbreds), age (3.5 to 14 years) and milk yield (liter/day) were evaluated as possible risk factors. The serum calcium, inorganic phosphorus, magnesium and glucose concentrations of the 220 dairy cows were determined by using imported commercial kits. Dairy cows with serum calcium concentrations ≤ 8.0 mg/dl and serum glucose ≤ 44.0 mg / dl with positive ketone tests but not showing any clinical signs were considered SCHC and SCK respectively.

**Results:** The overall prevalence of SCHC was 30.0%, of which 32.11% were recorded in HF × L, 15.0% in SH × L and 20.0% in JS × L cross-bred cows. The overall prevalence of SCK was 25.0%, of which 27.37% in HF × L, 10.0% in SH × L and 10.0% in JS × L cross-bred cows. The SCHC was recorded 10 times greater than MF and SCK 6 times greater than CK in Bangladesh. The hypocalcaemia and hypophosphatemia with hypermagnesemia status were recorded in SCHC affected lactating cows which were more significantly ( $p < 0.05$ ) higher (46.67%) at 4<sup>th</sup> parity and lower (16.67%) at 1<sup>st</sup> parity. The significantly ( $p < 0.01$ ) higher prevalence of SCK was recorded at the 4<sup>th</sup> (53.33%) in comparison to other parity especially lowest at 1<sup>st</sup> (2.78%) and 2<sup>nd</sup> (4.0%) parity. The significantly ( $p < 0.1$ ) highest prevalence of SCHC and SCK were recorded at high milk yield during the 1<sup>st</sup> (94.44%; 77.78%) and 2<sup>nd</sup> (66.67%; 56.67%) weeks of lactation period than the higher lactation stages respectively. The effects of BCS on the milk yield and the prevalence of SCHC and SCK are presented and discussed. Encouraging results with increased blood calcium and glucose levels were obtained on the therapeutic response of SCHC with oral calcium and SCK with oral propylene glycol.

**Conclusions:** The SCHC and SCK have detrimental effects on cow health, productivity and reproduction and also predisposes to other diseases and disorders. The efficient balanced ration, periodic screening blood, milk and urine for determination of concerned biochemical constituents and ketone bodies considering risk factors could help to early detection of SCHC and SCK to limit their effects in dairy cattle. The high prevalence of SCHC and SCK recorded in this study should be viewed as a potential health risk to the transition cows that requires further research.

**Keywords:** Sub-clinical milk fever, Sub-clinical ketosis, crossbred cows, prevalence, risk factors, calcium, magnesium, phosphorus, glucose, ketone bodies, therapeutic management, Bangladesh

Article Info: Article Code No. © LEP: JVMOHR/0020/2020

Received: 30 April 2020

Revised: 10 May 2020

Accepted: 11 June 2020

Published: 30 June 2020

**Citation:** Naher L, Samad MA, Siddiki SHMF and Islam MT (2020). Prevalence and risk factors of subclinical milk fever and ketosis in lactating cross-bred dairy cows with their therapeutic management in Bangladesh *J. Vet. Med. OH Res.* 2 (1): 139-182 [doi: 10.36111/jvmohr.2020.2 (1).0020]



**Copy right** © 2020. The Authors. Published by LEP. This is an open access article under the CC-BY-NC-ND License (<http://creativecommons.org/licenses/by-nc-nd/4.0/>)

## INTRODUCTION

Bangladesh is a tropical country with a total of 24.086 million cattle population which includes 80 to 90% indigenous and 10 to 15% cross-bred cattle,<sup>1-3</sup> of which 3.53 million lactating and 2.61 million dry cows.<sup>4</sup> In the last two decades, a heavy demand of milk due to high rate of urbanization of people in Bangladesh have initiated to establish a large number of mini dairy farms with mainly high yielding crossbred cows throughout the country. Currently, there are approximately 58,590 DLS registered mini dairy farms in Bangladesh.<sup>5</sup> Most of these private dairy farms in Bangladesh are small in size with 73% contain less than 11 cows and 17% has 11 to 20 cows. Of these dairy farms, 65% are reared by stall feeding system, 30% by stall cum open feeding system and the rest 5% by open feeding system.<sup>6</sup> The milk production per lactation is ranged from 300 to 400 liter in indigenous cows and 600 to 800 liter in cross-bred cows.<sup>7</sup> The dairy cattle in Bangladesh generally consist of indigenous<sup>8</sup> and crossbreds including mostly Sindhi, Sahiwal, Holstein Friesian and Jersey cross cows.<sup>9,10</sup> An indigenous cow has been reported to produce an average of 1.91 liter milk whereas cross-bred cow produces 7.80 liter milk per day at the smallholder farmers' management system in Bangladesh.<sup>1</sup> An overall 2,84,500 smallholder milk suppliers are being supplying milk to the 14 milk processing and marketing organizations in Bangladesh.<sup>11</sup> The dairy industry in Bangladesh has made tremendous strides in improving the average milk production per cow during the last two decades, mainly by the improvement of the genetic pool of indigenous cattle through cross-breeding program by using AI. The genetic progressed of cows have been made but the availability of feeds and fodder with feed efficiency has not yet been progressed at the same rate as milk production in Bangladesh. Nutritional deficiencies, imbalances ration or erratic management of feeding programs for dairy cows might have already created various types of health problems especially metabolic diseases.<sup>12</sup> Prevalence of metabolic diseases is closely related to ration, dairy farm management and some extent to animal's genetics. Imbalance and inadequate feeding of high milk yielding dairy cattle at pregnancy and pre-partum are usually associated with marked metabolic abnormalities at transition period (3 weeks around parturition) that makes them more susceptible to metabolic and even infectious diseases.<sup>13,14</sup> The occurrence of metabolic disorders in dairy cows depend on the ability of the cows to cope with the metabolic demands of high milk production and its etiology can be traced back to insults that occur during transition period. An increased energy and calcium demands for colostrum and milk production, combined with a decline of dry matter intake (DMI) around parturition, can result NEB, increased lipid mobilization<sup>15,16</sup> and a reduction in blood concentrations of calcium.<sup>17,18</sup> These changes increase the risk of metabolic and infectious diseases with animal welfare concern and an important cause of production and economic losses to the dairy industry.<sup>19</sup> Susceptibility to infectious diseases at transitional period occurs due to immune suppression during the peri-parturient period.<sup>20-22</sup> The metabolic disorders are usually occur in the high milk yielding pure exotic and crossbred dairy animals and such population was very limited in the then East Pakistan and early Bangladesh and accordingly there is a dearth of research reports on the occurrence of these diseases in inland literature. However, an overall 2.97% Milk fever and 3.75% Ketosis cases in cows have been reported based on analysis of hospital clinical cases from Bangladesh.<sup>23</sup> Recently, an overall

25% prevalence of SCK<sup>24</sup> and application of metabolic profiles tests to detect the metabolic profiles in lactating cross-bred dairy cows have been evaluated in Bangladesh.<sup>10</sup> This paper describes the prevalence of sub-clinical milk fever (SCMF) and sub-clinical ketosis (SCK) with their associated risk factors and therapeutic management of these sub-clinical hypocalcemia and SCK in lactating dairy cross-bred cows of Bangladesh.

## MATERIALS AND METHODS

A cross-sectional study was conducted on randomly selected apparently healthy 220 cross-bred (190 Holstein Friesian = HF  $\times$  Local = L, 20 Sahiwal = SH  $\times$  Local = L and 10 Jersey = JS  $\times$  Local = L) lactating cows of nine dairy farms and one smallholders' farm, of which two located in the district of Mymensingh and eight in the district of Gazipur during the period from July to November 2016. Two farms of Mymensingh district include ① Bangladesh Agricultural University (BAU) Dairy Farm (n = 45) and ② Smallholder dairy farm of adjacent villages of BAU Campus (n = 20). Randomly selected eight dairy farms in the district of Gazipur include ① Dipti and Sons Farm House (DFH), Valkartek (n = 35), ② Zahir Dairy Farm (ZDF), Dhirashrom (n = 35), ③ Alim Dairy Farm (ALDF), Bolodha (n = 25), ④ Masum Dairy Farm (MDF), Amuna (n = 15), ⑤ Azafor Dairy Farm (AZDF), Pragao (n = 12), ⑥ Mominul Dairy Farm (MDF), Aturi (n = 11), ⑦ Jaman Dairy Farm (JDF), Dakshin Khan (n = 11) and ⑧ Apon Dairy Farm (APDF), Valkartek (n = 11). These randomly selected lactating cows aged between 3.5 to 14 years, at different lactation stages, parity and level of milk production. The animals of the selected dairy farms are reared under semi-intensive management system with raised floor. They are often provided with water hyacinth, maloncha, Jumbo grass, green grass in addition to concentrate diet and feeding two times daily. These dairy cattle are kept together in common shed but they are maintained in separate shed at transition period. The cross-breed dairy cows selected at the adjacent villages of BAU campus, Mymensingh are maintained under traditional rural husbandry practices.

A structured questionnaire was used to collect animal and farm level data on age, breed, parity, body condition score (BCS), previous milk production record, present milk production, lactation stages (weeks), number of lactating cows in the herd, status of calves, feeds and feedings including grazing, milking system, disease and treatment history of all the selected dairy farms. These data were collected by interviewing the farm owners and in some cases abstracting the farm records.

## Collection and testing of urine samples

Fresh urine samples from each of 220 randomly selected lactating dairy cows were collected conveniently with the help of farm attendants or owners in plastic sample containers and tested directly at cow-side level or taken to the laboratory for the determination of ketone body in the urine. Each of the collected urine samples was tested for the presence of ketone body in the urine by using urinalysis reagent strips Uric 10 CF (Atena Medical Instrument Co., Guangdong, China). The urinalysis reagent strips are plastic strips to which chemically specific reagent pads are affixed. The reagent pads react with the sample urine to provide a standardized visible color reaction within 30 seconds to 1 minute depending on the specific panel screen. The color is

then visually compared to the included color chart to determine the level of each chemical factor. Each of the 10 reactive reagent pads on the test strip was compared to the corresponding line of color blocks on the chart. The closest color match indicated the test result. The test procedure was used as per instruction of the kit manufacturer company. In briefly, after collection of the urine samples in a clean dry plastic container, one reagent strip was removed from the bottle and immediately the container cap was replaced minimizing the exposure of the remaining test strips to light and air. The reagent pads of the strip were immersed completely in the urine sample and then removed immediately to avoid dissolving out the reagent pads. While removing the reagent strip, the edge of the strip was run against the rim of the specimen container to remove excess urine. The strip was then held in a horizontal position pads. The color change of the reagent pads was compared to the corresponding color chart on the bottle label. According to the chart's timeframe (i.e. 45 seconds for ketone body) the reading was taken. The colors range from beige or buff-pink color for a 'negative' reading to pink and pink-purple for a 'positive' reading.

#### **Collection and testing of serum samples**

About 10 ml of blood samples of each of the 220 lactating dairy cows were collected by using sterile disposable syringe and transferred into falcon tube without adding any anticoagulant and kept at room temperature for three hour. The blood samples were then kept in the refrigerator overnight at 4 °C. Then the blood samples were centrifuged at 3000 rpm for 15 minutes and serum was collected in Eppendorf tube by using pasture pipette and stored at - 20 °C until analysis.

The selective biochemical parameters e.g. serum calcium, phosphorus, magnesium and glucose levels were determined by using commercial test kits as per instruction of the kit manufacturing companies at the Central Laboratory of the BAU, Mymensingh.<sup>10</sup> The serum calcium concentration was determined with quantitative colorimetric Kit Calcium Arsenazo III (Reactivous GPL, Barcelona, Spain), inorganic phosphorus by using quantitative colorimetric Kit Vitro Inorganic phosphorus reagent (In vitro Diagnostics, Vitro Scient, Egypt) and the magnesium concentration by using quantitative colorimetric Kit (Magnesium Xylidyl Blue, Prestige Diagnostics, UK). The serum glucose concentration was determined by glucose oxidase (GOD) and peroxidase (POD) method using enzymatic qualitative colorimetric kit LABKIT reagents (Glucose GOD-POD Liquid, Barcelona, Spain).

#### **Therapeutic management of SCHC and SCK affected cows**

A total of 66 SCHC and 55 SCK affected lactating cross-bred cows were selected for treatment trials. Each of the SCHC affected cow was treated with calcium bolus (CP-Vet Plus<sup>®</sup> bolus, The Acme Laboratories Ltd.) @ 4 bolus / animal orally once daily for 5 days. Each CP-Vet Bolus contains calcium 830 mg, phosphorus 500 mg, Magnesium 375 mg, Potassium 250 mg, Sodium 125 mg, Vitamin D<sub>3</sub> 12500 iu, Vitamin E 75 iu and Vitamin B<sub>12</sub> 25 mcg. Each of the SCK affected cows received propylene glycol (Vita-D Plus<sup>®</sup> The Acme Laboratories Ltd.) @ 200 ml / animal orally, administered after mixing with equal volume of water twice daily for first two days and then half of the dosage for next two day. Each 100 ml of Vita-D Plus

contains Vitamin D<sub>3</sub> 500000 iu and Propylene glycol q.s to 100 ml. The serum calcium and glucose levels were estimated at the pre-treatment and post-treatment of all the treated cows after 7 days to detect the effectiveness of the administered drugs.

### Statistical analysis

Data were entered in Microsoft Excel 2010 and transferred to IBM SPSS (Statistical Package for Social Science) statistics 20.0 software. Z test for comparison of proportion, Chi-square test, Paired 't' test, Odd ratio, 95% Confidence interval and p-value calculation were done to find out the significant differences in the prevalence of bovine SCMF and SCK in terms of breed, age, parity, BCS, lactation stage, milk yield and blood glucose level of cows.

## RESULTS

A total of 220 cross-bred (190 HF × L, 20 SH × L and 10 JS × L) lactating dairy cows of 10 dairy herds between 1 to 13 weeks of lactation with high producing records were tested for SCMF and SCK. The SCMF was diagnosed based on the hypocalcemia ( $\leq 8$  mg/dl) and SCK by the detection of positive level of urinary ketone bodies using 10 CF urinalysis reagent strip and hypoglycemia ( $\leq 44$  mg/dl).

### Prevalence and risk factors of SCMF

The overall 30.0% prevalence of SCMF (sub-clinical hypocalcemia = SCHC) was recorded in lactating dairy cows, of which highest prevalence was found in HF × L (32.11%), followed by JS × L (20.0%) and lowest in SH × L (15.0%) cross-bred cows (Table 1 & Fig 1). Serum calcium, inorganic phosphorus and magnesium concentrations were estimated in all the 220 lactating cows, of which 66 (30.0%) had both the hypocalcemia ( $\leq 8.0$  mg/dl) and hypophosphatemia ( $< 4.0$  mg/dl) but all these 66 (30.0%) hypocalcemic cows had hypermagnesemia ( $>3$  mg/dl). These hypocalcemia and hypophosphatemia with hypermagnesemia findings were higher in HF × L, followed by JS × L and HS × L cross-bred of lactating cows (Table 1 & Fig 1). The overall hypocalcemia and hypophosphatemia with hypermanesemia were found significantly ( $p < 0.05$ ) higher at  $> 5$  years of age in comparison to  $< 4$  years and 4 to 5 years of age groups in lactating dairy cows (Table 1 & Fig. 2).

The hypocalcemia and hypophosphatemia with hypermagnesemia status were also observed in all the parity from 1<sup>st</sup> to 8<sup>th</sup> but significantly ( $p < 0.05$ ) lower (16.67%) at 1<sup>st</sup> parity in comparison to other parity (Table 2 & Fig. 3).

Table 3 shows significantly ( $p < 0.01$ ) higher prevalence of SCMF (hypocalcemia) with hypophosphatemia and hypermagnesemia at the 1<sup>st</sup> week of lactation in comparison to 2<sup>nd</sup> to 13<sup>th</sup> weeks of lactation stages (Fig. 4). Table 4 presents the effects of milk production and BCS on the prevalence of SCHC and its relationship with blood phosphorus and magnesium concentrations in lactating dairy cross-bred cows. A strong relationship was observed between high milk production and the prevalence of SCHC in the lactating dairy cows. The lactating cows producing milk  $> 15$  liter / day had significantly higher ( $p < 0.05$ ) and 2.5% cows producing  $< 5$  liter milk / day had a significantly lower ( $p < 0.05$ ) prevalence of SCHC (Table 4 & Fig. 5).

Table 1. Breeds and age factors associated with the prevalence of sub-clinical hypocalcemia and its relationship with phosphorus and magnesium concentrations in lactating dairy crossbred cows

S/ N	Variable	No. of cows tested	Serum macro-mineral levels (mg/dl)					
			Calcium Range Mean ± SE	SCHC (≤ 8 mg/dl) <sup>1</sup> Positive No. (%)	Phosphorus Range Mean ± SE	HP (< 4mg/dl) <sup>1</sup> Positive No. (%)	Magnesium Range Mean ± SE	HM (>3 mg /dl) <sup>1</sup> Positive No. (%)
A. Breeds								
①	HF × L	190	06.05-11.98 09.06 ± 0.13	61 (32.11)	2.47 – 6.99 3.30 ± 0.08	61 (32.11)	1.43-4.25 2.59 ± 0.06	61 (32.11)
②	SH × L	020	06.32 - 11.68 10.29 ± 0.40	03 (15.00)	3.13 - 6.97 5.13 ± 0.25	03 (15.00)	1.71-3.67 2.18 ± 0.14	03 (15.00)
③	JS × L	010	06.13 - 11.36 09.38 ± 0.58	02 (20.00)	3.04 – 5.98 4.46 ± 0.31	02 (20.00)	1.72-3.86 2.4 ± 0.26	02 (20.00)
	Overall	220	06.05 -11.98 09.19 ± 0.12	66 (30.00)	2.47 – 6.99 4.38 ± 0.08	66 (30.00)	1.43-4.25 2.55 ± 0.06	66 (30.00)
B. Age (years)								
①	< 4.0	036	06.67 - 11.68 09.75 ± 0.25	06 (16.67)	2.86-6.82 4.76 ± 0.18	06 (16.67)	1.67-3.77 2.3 ± 0.09	06 (16.67)
②	4 – 5	075	06.13 - 11.98 09.74 ± 0.21	19 (25.33)	2.84-6.99 4.60 ± 0.13	19 (25.33)	1.56-3.98 2.34 ± 0.09	19 (25.33)
③	> 5	109	06.05 - 11.68 08.62 ± 0.17	41 (37.16)*	2.47-6.98 4.10 ± 0.10	41 (37.16)	1.43-4.25 2.77 ± 0.08	41 (37.16)
	Overall	220	06.05 - 11.98 09.19 ± 0.12	66 (30.00)	2.47 - 6.99 4.38 ± 0.08	66 (30.00)	1.43-4.25 2.55 ± 0.06	66 (30.00)
SCHC = Sub-clinical hypocalcemia      HP = Hypophosphatemia      HM = Hypermagnesemia *Significant at (p < 0.05) <sup>1</sup> Cut off points HF = Holstein Friesian      SH = Sahiwal      JS = Jersey      L = Local								

The higher prevalence of SCHC was recorded in lactating cows with higher BCS ( $> 3.5$ ) at 36% in comparison to lower BCS (3 to 3.25) at 25% (Table 4 & Fig. 6).

### Sub-clinical milk fever and ketosis in crossbred cows

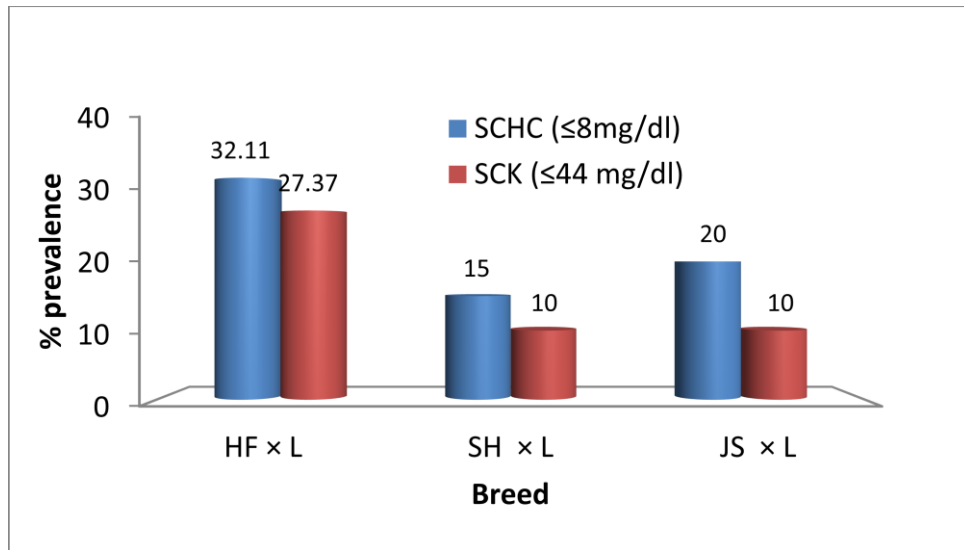


Fig.1. Breed-wise prevalence of subclinical hypocalcemia (SCHC) and subclinical Ketosis (SCK) in lactating cross-bred cows

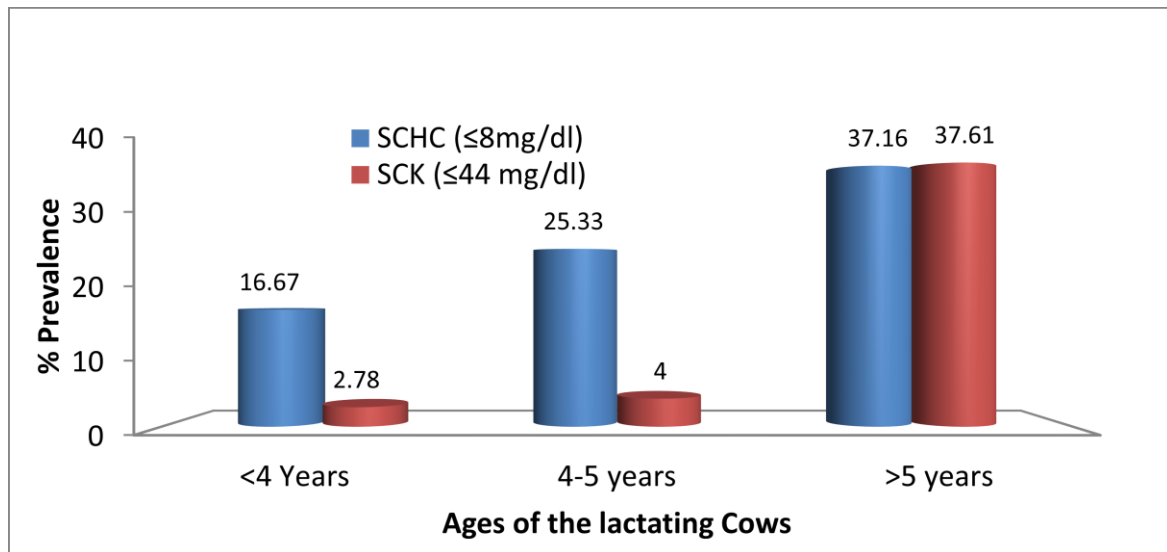


Fig.2. Age-wise prevalence of SCHC and SCK in lactating cross-bred cows



Table 2. Influence of parity on the prevalence of sub-clinical hypocalcemia and its relationship with phosphorus and magnesium in lactating dairy cross-bred cows							
Variable (Parity No.)	No. of cows tested	Serum macro-mineral levels (mg/dl)					
		Calcium Range Mean $\pm$ SE	SCHC ( $\leq 8$ mg/dl) <sup>1</sup> Positive No. (%)	Phosphorus Range Mean $\pm$ SE	HP ( $< 4$ mg/dl) <sup>1</sup> Positive No. (%)	Magnesium Range Mean $\pm$ SE	HM ( $>3$ mg /dl) <sup>1</sup> Positive No. (%)
1	36	6.67-11.68 9.75 $\pm$ 0.25	06 (16.67)	2.86-6.82 4.76 $\pm$ 0.18	06 (16.67)	1.67-3.77 2.3 $\pm$ 0.10	06 (16.67)
2	75	6.15-11.98 9.81 $\pm$ 0.21	19 (25.33)	2.84-6.99 4.63 $\pm$ 0.13	19 (25.33)	1.56-3.98 2.31 $\pm$ 0.84	19 (25.33)
3	40	6.09-11.68 9.05 $\pm$ 0.31	14 (35.00)	2.47-6.98 4.37 $\pm$ 0.21	14 (35.00)	1.43-4.25 2.64 $\pm$ 0.15	14 (35.00)
4	30	6.13-11.68 8.24 $\pm$ 0.29	14 (46.67)*	2.54-6.34 3.82 $\pm$ 0.15	14 (46.67)	1.65-4.25 2.96 $\pm$ 0.14	14 (46.67)
5	20	6.05-11.36 8.42 $\pm$ 0.37	07 (35.00)	3.11-5.87 3.96 $\pm$ 0.19	07 (35.00)	1.65-3.98 2.79 $\pm$ 0.19	07 (35.00)
6	11	6.21-10.77 8.09 $\pm$ 0.48	04 (36.36)	2.84-5.27 4.03 $\pm$ 0.25	04 (36.36)	1.72-3.67 2.69 $\pm$ 0.21	04 (36.36)
7	04	6.36-9.93 8.09 $\pm$ 0.86	01 (25.00)	2.71-4.32 3.5 $\pm$ 0.42	01 (25.00)	2.13-3.89 3.13 $\pm$ 0.47	01 (25.00)
8	04	6.0-11.68 8.91 $\pm$ 1.51	02 (25.00)	2.84-6.13 4.36 $\pm$ 0.79	01 (25.00)	1.68-3.94 2.78 $\pm$ 0.62	01 (25.00)
Overall	220	6.05-11.98 9.19 $\pm$ 0.12	66 (30.0)	2.47-6.99 4.38 $\pm$ 0.08	66 (30.00)	1.43-4.25 2.55 $\pm$ 0.06	66 (30.00)
*Significant at (p < 0.05)				<sup>1</sup> Cut off points			



# Sub-clinical milk fever and ketosis in crossbred cows

Table 3. Influence of lactation stages on the prevalence of sub-clinical hypocalcemia and its relationship with phosphorus and magnesium in lactating cross-bred cows

Variable (Lactation stages: weeks)	No. of cows tested	Serum macro-mineral levels (mg/dl)					
		Calcium Range Mean $\pm$ SE	SCHC ( $\leq 8$ mg/dl) <sup>1</sup> Positive No. (%)	Phosphorus Range Mean $\pm$ SE	HP ( $< 4$ mg/dl) <sup>1</sup> Positive No. (%)	Magnesium Range Mean $\pm$ SE	HM ( $>3$ mg/dl) <sup>1</sup> Positive No. (%)
1	18	06.13-07.6 06.68 $\pm$ 0.13	17 (94.44)**	2.47-4.15 2.94 $\pm$ 0.10	17 (94.44)	3.11-4.25 3.87 $\pm$ 0.07	17 (94.44)
2	30	06.05-11.36 07.54 $\pm$ 0.25	20 (66.67)	2.54-6.12 3.63 $\pm$ 0.15	20 (66.67)	1.71-4.25 3.19 $\pm$ 0.13	20 (66.67)
3	35	06.20-11.68 08.91 $\pm$ 0.25	10 (28.57)	2.84-6.34 4.2 $\pm$ 0.14	10 (28.57)	1.71-3.94 2.66 $\pm$ 0.12	10 (28.57)
4	40	06.45-11.68 09.16 $\pm$ 0.22	09 (22.50)	2.84-6.34 4.27 $\pm$ 0.13	09 (22.50)	1.43-3.58 2.43 $\pm$ 0.10	09 (22.50)
5	25	07.13-11.36 09.11 $\pm$ 0.21	04 (16.00)	2.94-6.56 4.25 $\pm$ 0.16	04 (16.00)	1.56-3.54 2.42 $\pm$ 0.12	04 (16.00)
6	20	06.22-11.68 09.83 $\pm$ 0.32	03 (15.00)	3.23-6.98 4.61 $\pm$ 0.22	03 (15.00)	1.67-3.54 2.13 $\pm$ 0.12	03 (15.00)
7	10	07.23-11.68 10.31 $\pm$ 0.61	02 (20.00)	2.84-6.88 5.24 $\pm$ 0.43	02 (20.00)	1.65-3.87 2.21 $\pm$ 0.26	02 (20.00)
8	08	09.13-11.68 10.94 $\pm$ 0.38	0	4.16-6.93 5.77 $\pm$ 0.41	0	1.71-2.21 1.87 $\pm$ 0.07	0
9	09	06.39-11.68 10.49 $\pm$ 0.65	01 (11.11)	3.22-6.49 5.29 $\pm$ 0.37	01 (11.11)	1.71-3.78 2.16 $\pm$ 0.25	01 (11.11)
10	08	09.23-11.98 11.34 $\pm$ 0.31	0	4.01-6.23 5.23 $\pm$ 0.21	0	1.69-2.54 1.88 $\pm$ 0.10	0
11	08	10.72-11.68 11.56 $\pm$ 0.12	0	5.27-6.99 5.77 $\pm$ 0.27	0	1.71-1.79 1.76 $\pm$ 0.01	0
12	08	11.36-11.68 11.60 $\pm$ 0.05	0	5.27-6.33 5.65 $\pm$ 0.19	0	1.71-1.79 1.77 $\pm$ 0.01	0
13	01	11.68	0	6.54	0	1.79	0

\*\*Significant at (p < 0.01)

<sup>1</sup>Cut off values

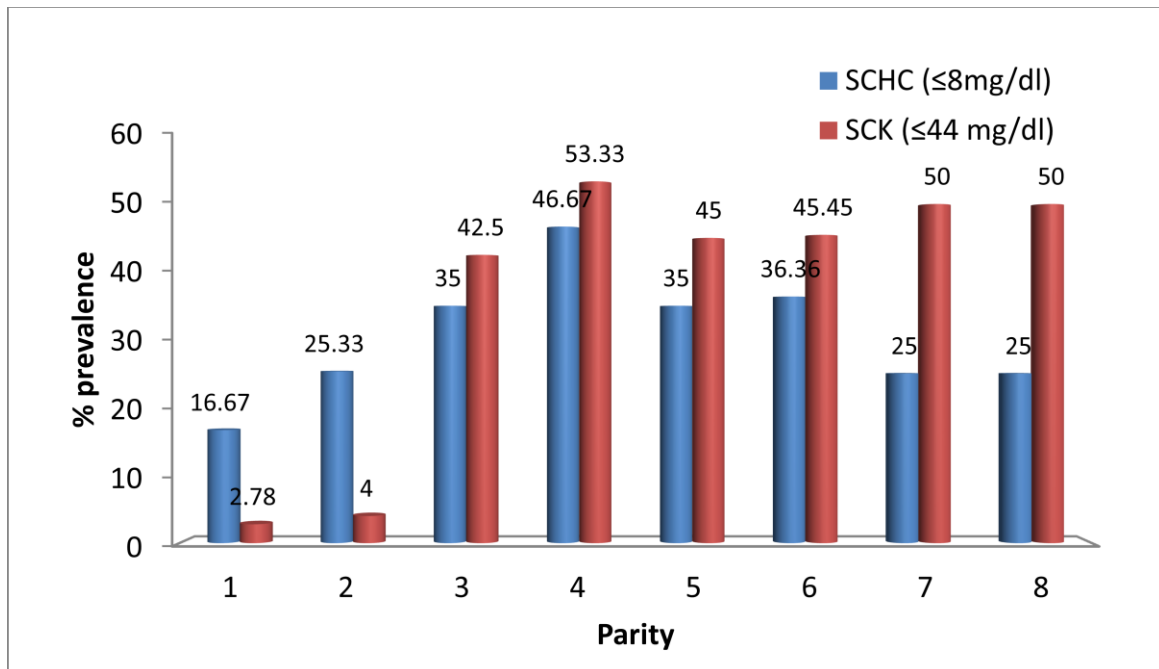


Fig. 3. Parity-wise prevalence of SCHC and SCK in lactating cross-bred dairy cows

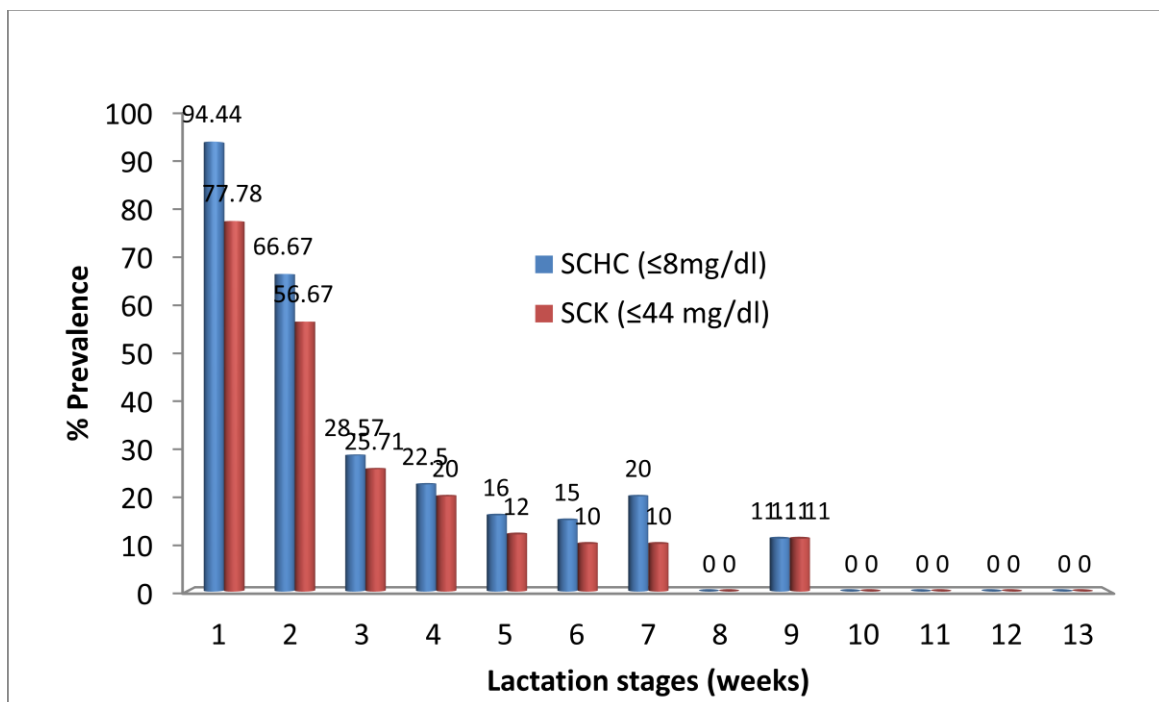


Fig. 4. Lactation stage-wise prevalence of SCHC and SCK in lactating dairy cows

## Sub-clinical milk fever and ketosis in crossbred cows

Table 4. Effects of milk yield (liter / day) and body condition score (BCS) on the prevalence of subclinical hypocalcemia and its relationship with phosphorus and magnesium in lactating cross-bred cows							
S/ Variables N	No. of cows tested	Serum macro-mineral levels (mg/dl)					
		Calcium Range Mean ± SE	SCHC (≤ 8 mg/dl) <sup>1</sup> Positive No. (%)	Phosphorus Range Mean ± SE	HP ( $< 4$ mg/dl) <sup>1</sup> Positive No. (%)	Magnesium Range Mean ± SE	HM ( $>3$ mg /dl) <sup>1</sup> Positive No. (%)
① Milk yield (liter/day)							
< 05.0	040	07.89-11.68 11.38 ± 0.13	01 (2.50)**	3.81-6.99 5.7 ± 0.12	01 (2.50)	1.65-3.10 1.83 ± 0.05	1 (2.50)
05.0-10.0	030	07.23-11.98 10.28 ± 0.31	06 (20.00)	2.94-6.98 5.20 ± 0.21	06 (20.00)	1.65-3.23 2.17 ± 0.11	06 (20.00)
11.0-15.0	115	06.13-11.68 08.66 ± 0.13	35 (30.43)	2.84-6.34 4.03 ± 0.07	35 (30.43)	1.43-3.98 2.63 ± 0.07	35 (30.43)
> 15.0	035	06.05-09.93 07.47 ± 0.23	24 (68.57)*	2.47-4.76 3.32 ± 0.11	24 (68.57)	2.13-4.25 3.43 ± 0.13	24 (68.57)
Overall	220	06.05-11.98 09.19 ± 0.12	66 (30.00)	2.47-6.99 4.38 ± 0.08	66 (30.0)	1.43-4.25 2.55 ± 0.06	66 (30.00)
② Body condition score (BCS)							
3.0-3.25	120	06.13-11.98 09.41 ± 0.16	30 (25.00)	2.84-6.99 4.54 ± 0.09	30 (25.00)	1.56-3.98 2.46 ± 0.07	30 (25.00)
> 3.5	100	06.05-11.68 08.92 ± 0.19	36 (36.00)	2.47-6.98 4.20 ± 0.11	36 (36.00)	1.43-4.25 2.66 ± 0.09	36 (36.00)
Overall	220	06.05-11.98 09.19 ± 0.12	66 (30.00)	2.47-6.99 4.38 ± 0.08	66 (30.00)	1.43-4.25 2.55 ± 0.06	66
*Significantly higher at (p < 0.05)			**Significantly lower at (p < 0.05)			<sup>1</sup> Cut off points	

The potential risk factors for SCHC in lactating dairy cows associated with breed, ages, parity, lactation stages, milk yield and BCS have been analyzed and evaluated (Table 5).

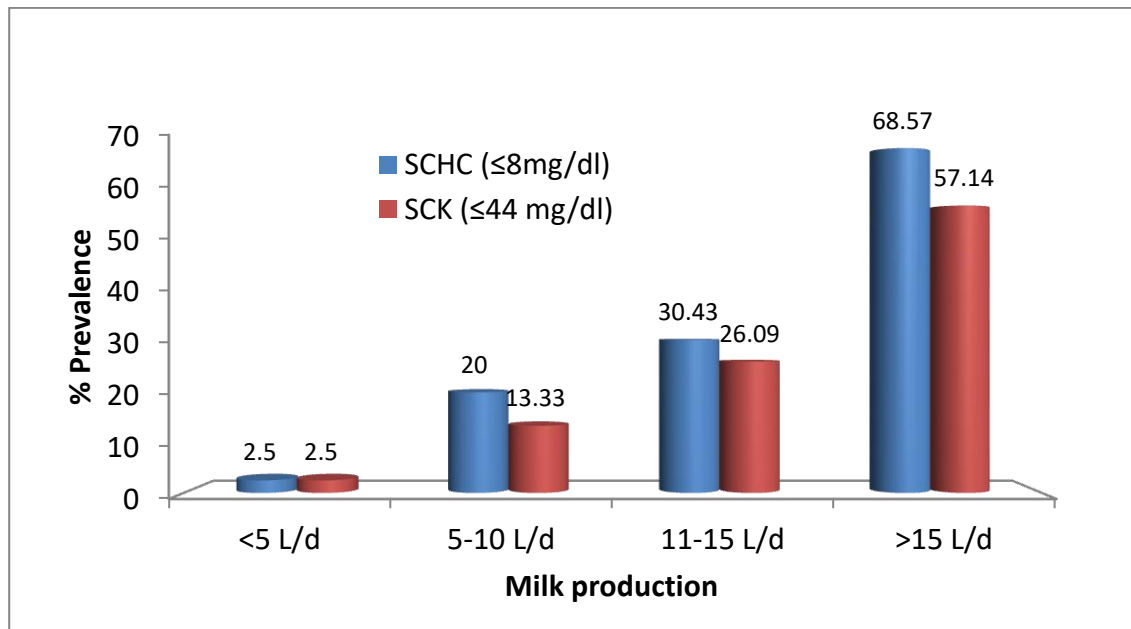


Fig. 5. Milk production-wise prevalence of SCHC and SCK in lactating dairy cows

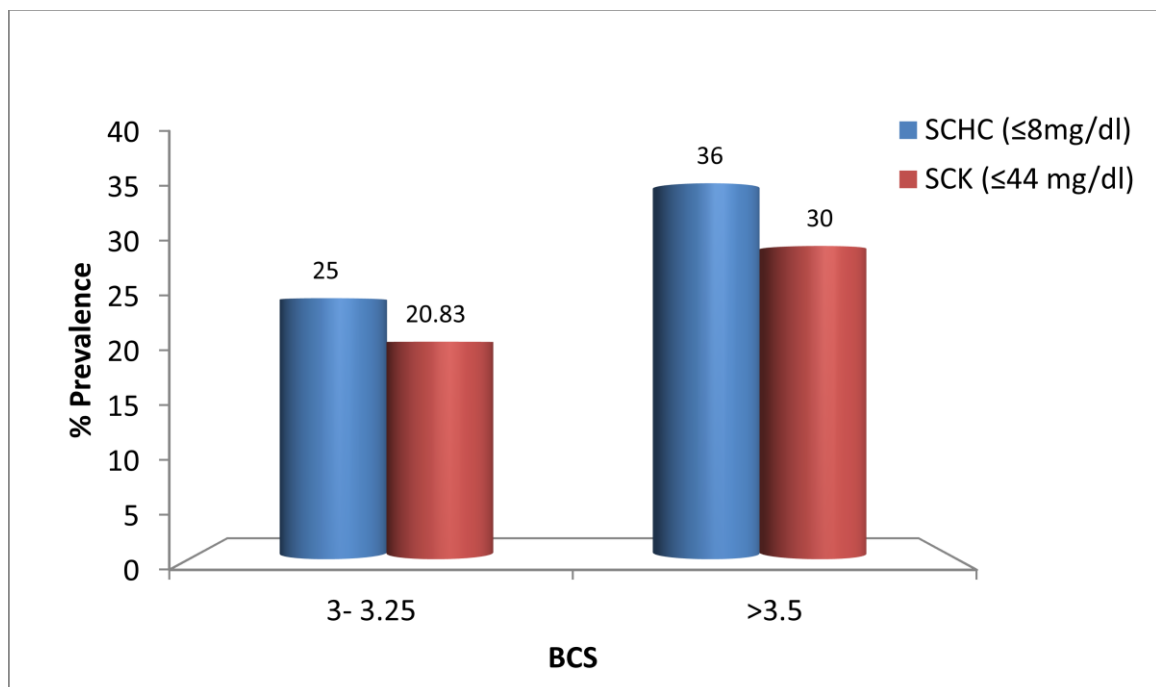


Fig. 6. Body condition score (BCS)-wise prevalence of SCHC and SCK in lactating cows

# Sub-clinical milk fever and ketosis in crossbred cows

Table 5. Risk factors analysis of subclinical hypocalcemia (SCHC) in lactating crossbred dairy cows

S/N	Risk factors	Categories	SCHC		Odd ratio	95% CI	p-value
			Positive (n = 66)	Negative (n = 154)			
1. Breed		HF × L	61	129	02.68	00.76 - 09.49	0.13
		SH × L	03	017	Reference		
		JS × L	02	008	01.42	00.20 - 10.23	0.73
2. Ages (years)		< 4.0	06	030	Reference		
		4.0-50	19	056	01.70	00.61 - 04.70	0.31
		> 5.0	41	068	03.02	01.16 - 07.86	0.02
3. Parity		01	06	030	Reference		
		02	19	056	01.70	00.61 - 04.70	0.31
		03	14	026	02.69	00.90 - 08.02	0.08
		04	14	016	04.38	01.41 - 13.58	0.01
		05	07	013	02.69	00.76 - 09.59	0.13
		06	04	007	02.86	00.63 - 12.92	0.17
		07	01	003	01.67	00.15 - 18.88	0.68
		08	01	003	01.67	00.15 - 18.88	0.68
4. Lactation stages (week)		01	17	001	136.00	07.51 - 2463	0.0009
		02	20	010	16.00	01.75 - 146.3	0.01
		03	10	025	03.20	00.35 - 29.01	0.30
		04	09	031	02.32	00.26 - 21.12	0.45
		05	04	021	01.52	00.15 - 15.78	0.72
		06	03	017	01.41	00.13 - 15.78	0.78
		07	02	008	02.00	00.15 - 26.74	0.60
		08	00	008	-	-	-
		09	01	008	Reference		
		10	00	008	-	-	-
		11	00	008	-	-	-
		12	00	008	-	-	-
		13	00	001	-	-	-
5. Milk yield (liter/day)		< 5.0	01	039	Reference		
		05-10	06	024	09.75	01.11 - 86.02	0.04
		10-15	35	080	17.06	02.25 - 129.2	0.006
		> 15.0	24	011	85.09	10.30 - 701.4	< 0.0001
6. Body condition score (BCS)		03-3.25	30	090	Reference		
		> 3.5	36	064	01.69	00.94 - 03.02	0.08
95% CI = 95% Confidence interval							

### Prevalence and risk factors of SCK

Out of 220 lactating dairy cows tested, of which 55 (25.0%) animals had SCK detected by the both hypoglycemia and urinary ketone bodies tests (Table 6). The highest prevalence of SCK was recorded in HF x L (27.37%) in comparison to SH x L (10.0%) and JS x L (10.0%) and > 5 years of age had significantly ( $p < 0.01$ ) higher prevalence (46.78%) in comparison to < 4 years (2.78%) and 4 to 5 years (4.0%) age groups in lactating cows (Table 6 & Fig. 1-2).

Table 6. Breeds and age-wise prevalence of sub-clinical ketosis detected by blood glucose and urinary ketone bodies in lactating dairy crossbred cows						
S/ Variables N	No. of cows tested	Serum glucose level (mg/dl)				Urinary ketone bodies test
		Range Mean $\pm$ SE	Sub-clinical ketosis		Total	Positive No. (%)
			$\leq 35$ mg <sup>1</sup> Positive No. (%)	36-44 mg <sup>1</sup> Positive No. (%)		
A. Breed						
① HF $\times$ L	190	20.83-86.54 55.02 $\pm$ 1.42	43 (22.63)	09 (04.74)	52 (27.37)	52 (27.37)
② SH $\times$ L	020	26.04-86.54 64.66 $\pm$ 4.17	02 (10.00)	0	02 (10.00)	02 (10.00)
③ JS $\times$ L	010	41.66-83.67 60.35 $\pm$ 4.85	0	01 (10.00)	01 (10.00)	01 (10.00)
Overall	220	20.83-86.54 56.14 $\pm$ 1.32	45 (20.45)	10 (04.55)	55 (25.00)	55 (25.00)
B. Age (years)						
① < 4.0	036	36.45-86.54 62.09 $\pm$ 2.16	01 (02.78)	0	01 (02.78)	01 (02.78)
② 4.0-5.0	075	20.83-86.54	03 (04.00)	0	03 (04.00)	03 (04.00)
③ > 5.0	109	20.83-86.54 48.21 $\pm$ 2.03	41 (37.61)	10 (09.17)	51 (46.78)**	51 (46.78)**
Overall	220	20.83-86.54 56.14 $\pm$ 1.32	45 (20.45)	10 (04.55)	55 (25.00)	55 (25.00)
**Significant at p < 0.01) <sup>1</sup> Cut off points HF = Holstein Friesian SH = Sahiwal JS = Jersey $\leq 35$ mg/dl considered CK <sup>108</sup>						

The influence of parity on the prevalence of SCK in lactating crossbred cows was recorded through the significantly ( $p < 0.01$ ) highest prevalence of SCK occurred during the 4<sup>th</sup> (53.33%) parity and significantly ( $p < 0.01$ ) lower prevalence at the 1<sup>st</sup> (2.78% and 2<sup>nd</sup> (4.0%) parity (Table 7 & Fig. 3).

## Sub-clinical milk fever and ketosis in crossbred cows

Table 7. Influence of parity on the prevalence of sub-clinical ketosis detected by blood glucose and urinary ketone bodies in lactating dairy crossbred cows

Variables (Parity No.)	No. of cows tested	Serum glucose level (mg/dl)				Urinary ketone bodies test
		Range (Mean $\pm$ SE)	Sub-clinical ketosis		Total	Positive No. (%)
			$\leq 35$ mg Positive No. (%)	36-44 mg Positive No. (%)		
1	36	36.45-86.54 (62.09 $\pm$ 2.16)	01 (02.78)	0	01 (02.78)**	01 (02.78)
2	75	20.83-86.54 (65.09 $\pm$ 1.65)	03 (04.00)	0	03 (04.00)**	03 (04.00)
3	40	20.83-86.54 (52.34 $\pm$ 3.52)	13 (32.50)	04 (10.00)	17 (42.50)	17 (42.50)
4	30	20.83-69.27 (44.65 $\pm$ 3.75)	13 (43.33)	03 (10.00)	16 (53.33)*	16 (53.33)*
5	20	20.83-76.56 (48.87 $\pm$ 4.41)	06 (30.00)	03 (15.00)	09 (45.00)	09 (45.00)
6	11	26.04-76.56 (43.08 $\pm$ 5.04)	05 (45.45)	0	05 (45.45)	05 (45.45)
7	04	20.83-52.08 (36.45 $\pm$ 7.67)	02 (50.00)	0	02 (50.00)	02 (50.00)
8	04	20.83-83.67 (50.65 $\pm$ 17.26)	02 (50.00)	0	02 (50.00)	02 (50.00)
Overall	220	20.83-86.54 (56.14 $\pm$ 1.32)	45 (20.45)	10 (04.55)	55 (25.00)	55 (25.00)
*Significantly higher at (p < 0.01)      ** Significantly lower at (p < 0.01) <sup>1</sup> Cut off point						

The prevalence of SCK was recorded from 1<sup>st</sup> to 9<sup>th</sup> weeks of lactation period but significantly ( $p < 0.01$ ) highest prevalence was recorded at the 1<sup>st</sup> (77.778%) and 2<sup>nd</sup> (56.67%) weeks of lactation period than the higher lactation weeks (Table 8 & Fig. 4). Table 9 shows the effects of milk production and BCS on the prevalence of SCK in dairy lactating cows. It appears that the higher milk production is associated with the higher prevalence of SCK and a significantly ( $p < 0.01$ ) higher prevalence of SCK (57.14%) was recorded in the cows producing > 15 liter milk / day (Table 9 & Fig.5). The higher prevalence of SCK (30.0%) was observed with higher BCS (>3.5) than 20.83% with lower BCS (3.0 - 3.25) in lactating dairy crossbred cows (Table 9 & Fig. 6).

The potential risk factors for sub-clinical ketosis in lactating dairy cows associated with breed, ages, parity, lactation stages, milk yield and BCS have been analyzed and evaluated (Table 10).



Table 8. Influence of lactation stages on the prevalence of sub-clinical ketosis detected by blood glucose and urinary ketone bodies in lactating dairy crossbred cows

Variables (Lactation stages (weeks))	No. of cows tested	Serum glucose level (mg/dl)		Sub-clinical ketosis			Urinary ketone bodies test	
		Range (Mean $\pm$ SE)	$\leq 35$ mg <sup>1</sup> Positive No. (%)	36-44 mg <sup>1</sup> Positive No. (%)	Total Positive No. (%)		Positive No. (%)	
01	18	20.83-76.56 (32.31 $\pm$ 3.90)	13 (72.22)	01 (05.56)	14 (77.78)*		14 (77.78)	
02	30	20.83-76.56 (35.36 $\pm$ 2.87)	17 (56.67)	0	17 (56.67)*		17 (56.67)	
03	35	20.83-76.56 (50.74 $\pm$ 2.69)	08 (22.85)	01 (02.86)	09 (25.71)		09 (25.71)	
04	40	20.83-86.54 (56.90 $\pm$ 2.20)	05 (12.50)	03 (07.50)	08 (20.00)		08 (20.00)	
05	25	26.04-69.27 (61.27 $\pm$ 2.26)	01 (04.00)	02 (08.00)	03 (12.00)		03 (12.00)	
06	20	46.87-83.25 (65.62 $\pm$ 2.19)	0	02 (10.00)	02 (10.00)		02 (10.00)	
07	10	43.66-85.25 (71.02 $\pm$ 4.68)	0	01 (10.00)	01 (10.00)		01 (10.00)	
08	08	41.66-44.88 (74.37 $\pm$ 5.44)	0	0	0		0	
09	09	20.83-83.25 (68.74 $\pm$ 6.29)	01 (11.11)	01 (11.11)	01 (11.11)		01 (11.11)	
10	08	46.87-76.56 (70.12 $\pm$ 3.56)	0	0	0		0	
11	08	69.27-86.54 (80.23 $\pm$ 2.27)	0	0	0		0	
12	08	69.27-86.54 (75.99 $\pm$ 1.91)	0	0	0		0	
13	01	76.56	0	0	0		0	
Overall	220	20.83-86.54 (56.14 $\pm$ 1.32)	45 (20.45)	10 (04.55)	55 (25.00)		55 (25.00)	
*Significantly higher at (p < 0.01)			<sup>1</sup> Cut off points $\leq 35$ mg /dl considered CK <sup>108</sup>					

### Concurrent prevalence of sub-clinical hypocalcaemia and sub-clinical ketosis

Of the 220 lactating cross-bred dairy cows tested, of which 46 (20.91%) were affected with both the SCHC and SCK concurrently and only 09 (4.09%) and 20 (9.09%) affected alone with SCK and SCHC, respectively (Table 11). The 20.19% concurrent prevalence of SCHC and SCK was found significantly (p<0.001) higher in comparison to single occurrence of SCK (4.09%) and SCHC (9.09%) in lactating cross-bred cows (Table 11).

## Sub-clinical milk fever and ketosis in crossbred cows

Table 9. Milk yield (liter/day) and body condition score (BCS) associated with the prevalence of sub-clinical ketosis in lactating dairy crossbred cows

Variables	No. of cows tested	Serum glucose level (mg/dl)				Urinary ketone bodies test
		Range (Mean ± SE)	Sub-clinical ketosis			
			≤ 35 mg <sup>1</sup> Positive No. (%)	36-44 mg <sup>1</sup> Positive No. (%)	Total Positive No. (%)	
① Milk yield (liter/day)						
< 5.0	040	46.87-86.54 (76.88 ± 1.22)	01 (02.50)	0	01 (02.50)	01 (02.50)
5.0-10	030	36.45-86.54 (68.23 ± 2.50)	03 (10.00)	01 (03.33)	04 (13.33)	04 (13.33)
11-15	115	20.83-76.56 (52.25 ± 1.49)	22 (19.13)	08 (06.67)	30 (26.09)	30 (26.09)
>15.0	035	20.83-63.13 (34.83 ± 2.46)	20 (57.14)	0	20 (57.14)**	20 (57.14)
Overall	220	20.83-86.54 (56.14 ± 1.32)	45 (20.45)	10 (04.55)	55 (25.00)	55 (25.00)
② BCS						
3.0-3.25	120	20.83-86.54 (60.32 ± 1.56)	15 (12.50)	10 (08.33)	25 (20.83)	25 (20.83)
> 3.5	100	20.83-86.54 (51.10 ± 2.11)	30 (30.00)	0	30 (30.00)	30 (30.00)
Overall	220	20.83-86.54 (56.14 ± 1.32)	45 (20.45)	10 (04.55)	55 (25.00)	55 (25.00)
**Significantly higher at (p < 0.01)			<sup>1</sup> Cut off points	≤ 35 mg/dl considered CK <sup>108</sup>		

### Therapeutic management of SCHC and SCK affected cows

An overall blood calcium and glucose levels of both the SCHC and SCK affected lactating cows treated with CP-Vet bolus<sup>®</sup> (Calcium contained mineral and vitamin preparation bolus) and Vita-D Plus<sup>®</sup> (contained propylene glycol with vitamin D) increased significantly (p < 0.001) after seven days of post-treatment in comparison to pre-treatment values (Table 12) and Vita-D Plus<sup>®</sup> (contained propylene glycol with vitamin D) increased significantly (p < 0.001) after seven days of post-treatment in comparison to pre-treatment values (Table 12).

Table 10. Risk factors analysis of subclinical ketosis (SCK) in lactating crossbred cows						
S/N Risk factors	Categories	SCK		Odd ratio	95% CI	p-value
		Negative (n = 55)	Positive (n = 165)			
1. Breed	HF × L	52	138	03.39	00.42 - 27.43	0.25
	SH × L	02	018	01.00	00.08 - 12.56	1.00
	JS × L	01	009	Reference		
2. Ages (years)	< 4.0	01	035	Reference		
	4.0-50	03	072	01.46	00.15 - 14.53	0.75
	> 5.0	51	058	30.78	04.07 - 232.7	0.0009
3. Parity	01	01	035	Reference		
	02	03	072	01.46	00.15 - 14.53	0.75
	03	17	023	25.87	03.22 - 207.9	0.002
	04	16	014	40.00	04.83 - 332.1	0.0006
	05	09	011	28.64	02.26 - 251.9	0.002
	06	05	006	29.17	02.88 - 295.4	0.004
	07	02	002	35.00	02.15 - 570.8	0.01
	08	02	002	35.00	02.15 - 570.8	0.01
4. Lactation stages (week)	01	14	004	31.50	03.02 - 328.9	0.004
	02	17	013	11.77	01.32 - 105	0.027
	03	09	026	03.12	00.35 - 28.14	0.31
	04	08	032	02.25	00.25 - 20.44	0.47
	05	03	022	01.23	00.11 - 13.43	0.87
	06	02	018	01.00	00.08 - 12.56	1.00
	07	01	009	Reference		
	08	00	008	-	-	-
	09	01	008	01.125	00.06 - 21.09	0.94
	10	00	008	-	-	-
	11	00	008	-	-	-
	12	00	008	-	-	-
	13	00	008	-	-	-
5. Milk yield (liter/day)	< 5.0	01	039	Reference		
	05-10	04	026	06.00	00.63 - 56.75	0.12
	10-15	30	085	13.76	01.81 - 104.6	0.01
	> 15.0	20	015	52.00	06.40 - 422.4	0.0002
6. Body condition score (BCS)	03-3.25	25	095	Reference		
	> 3.5	30	070	01.63	00.88 - 03.01	0.12
95% CI = 95% Confidence interval						

## Sub-clinical milk fever and ketosis in crossbred cows

Table 11. Concurrent occurrence of sub-clinical ketosis (SCK) and sub-clinical hypocalcaemia (SCHC) in lactating dairy cows

SN	Cross-bred cows	No. of cows tested	SCK + SCHC Positive No. (%)	SCK Positive No. (%)	SCHC Positive No. (%)	Total Positive No. (%)
①	Holstein-Friesian × Local	190	44 (23.16)	08 (04.22)	17 (08.95)	69 (36.32)
②	Sahiwal × Local	020	02 (10.00)	0	01 (05.00)	03 (15.00)
③	Jersey × Local	010	0	01 (10.00)	02 (20.00)	03 (30.00)
	Overall	220	46 (20.91)**	09 (04.09)	20 (09.09)	75 (34.09)
**Significantly higher at (p < 0.001)						

Table 12. Comparison of blood calcium and glucose levels between pre and post-treatment of sub-clinical hypocalcemia and sub-clinical ketosis affected lactating dairy cows [Range, Mean ± SE]

SN	Cross-bred cows	Serum calcium levels (mg/dl)			Serum glucose levels (mg/dl)		
		No. of positive cows treated	Pre-treatment Range Mean ± SE	Post-treatment Range Mean ± SE	No. of positive cows treated	Pre-treatment Range Mean ± SE	Post-treatment Range Mean ± SE
①	HF × Local	61	6.05-7.96 6.93 ± 0.07	09.56-12.87 11.58 ± 0.14	52	20.83-44.00 27.75 ± 1.13	45.27-59.45 53.08 ± 0.66
②	SH × Local	03	6.32-7.13 6.76 ± 0.24	10.34-11.23 10.93 ± 0.30	02	26.04-31.25 28.65 ± 2.61	45.27-58.34 51.81 ± 6.54
③	JS × Local	02	6.13-7.24 6.69 ± 0.56	12.21-12.87 12.54 ± 0.33	01	42.01	58.12
	Overall	66	6.05-7.96 6.92 ± 0.07	09.56-12.87 11.58 ± 0.13**	55	20.83-44.00 28.04 ± 1.10	45.27-59.45 53.12 ± 0.65**
**Significantly higher at (p < 0.01)		HF = Holstein-Friesian		SH = Sahiwal		JS = Jersey	

## DISCUSSION

Metabolic diseases are the multifactorial disorders of high milk yielding dairy cattle, primarily caused by imbalance and inadequate feeding, and erratic management of animals associated with heavy economic losses in dairy industry worldwide.<sup>12,19,25-27</sup> Approximately 75% of diseases in dairy cattle occur in the first month post-partum and 50% of dairy cattle suffer from metabolic and infectious diseases in the transition period.<sup>28</sup> The transition period which is the beginning at the last three weeks of pregnancy and extending into the third week of lactation that is around 3 weeks peri-parturition in cows.<sup>10,20,21</sup> The transition period during parturition and the initiation of lactation is very critical for the dairy cow production cycle because there are increased hormonal and metabolic changes.<sup>29</sup> There is an increased energy and calcium demands for colostrum and milk production, combined with a decline of dry matter intake (DMI) around parturition, can result NEB, increased lipid mobilization<sup>16,30</sup> and a reduction in blood concentrations of calcium.<sup>17,18</sup> This metabolic stress of the transition period of a dairy cow lead to a high incidence of metabolic, infectious and reproductive disorders associated with a severe negative energy balance (NEB). This period of NEB lasts approximately five weeks after calving.<sup>31</sup> Among the metabolic diseases, milk fever (MF) and ketosis occur commonly associated with transition period and peak milk production in dairy cattle.<sup>32</sup> Both the clinical and sub-clinical forms of metabolic diseases occur in dairy animals. However, the severity of the disease is higher in the clinical cases but subclinical cases are more important because (a) they are far more frequent, (b) they cannot be easily diagnosed, and (c) may impair the longevity and production of the cow.<sup>17,33</sup>

Dairy cows affected with clinical MF and clinical ketosis (CK) could be diagnosed in the field on the basis of disease history, clinical findings and response to therapy. However, dairy cows with SCMF and SCK do not show any clinical symptoms but have a SCHC and sub-clinical hypoglycemia (SCHG) with the positive level of ketone bodies regarded as SCK, respectively. Thus the only way to know whether dairy cows are experiencing SCHC and SCK are to analyze blood for the concentrations of specific biochemical constituents. In addition, blood, milk and urine samples could also be used to detect the ketone bodies for the diagnosis of SCK in dairy cows.

## Hypocalcemia

Hypocalcemia is one of the most common metabolic disorders in dairy cattle, classified into two forms, clinical hypocalcemia (parturient paresis (PP) / MF) and SCHC. Clinical hypocalcemia (MF) is easily identified, treated and readily responds to therapy when treatments are initiated promptly, whereas the SCHC cases cannot be treated easily due to the absence of obvious clinical signs required for diagnosis. The SCHC has been defined as low calcium concentrations (serum Ca  $\leq$  8.0 mg/dl) without any clinical symptoms of MF in dairy animals. Hypocalcaemia in dairy cows can also be called the 'gateway disease' as it increases the risk of other diseases and disorders like decreased milk production, decrease immune function, slower uterine involution, delayed first ovulation after calving, reduced gastro-intestinal motility, increased risk of ketosis, decreased reproductive performance, retained placenta, mastitis, metritis, endometritis, displaced abomasum and increased risk of early removal from the herd.<sup>17,32,34-39</sup>

## Sub-clinical milk fever and ketosis in crossbred cows

Approximately 82% of the cows and 5.1 times greater incidence of displaced abomasum had serum calcium values  $\leq 2.0$  mmol/L ( $\leq 8.0$  mg/dl) in the first week after calving.<sup>35,40</sup> Hypocalcemia associated with decrease or loss of muscle tone in the uterus increased the incidence of dystocia, uterine prolapse and retained placenta in post-partum dairy animals.<sup>32,41,42</sup> MF affected cows have been reported to be 3 to 6 times more susceptible to dystocia than that of normal cows.<sup>34,41-43</sup>

Hypocalcaemia is associated with impaired immune function and diminished muscle contraction that lead to develop metritis in post-parturient dairy cows.<sup>33,34,44</sup> It has also been reported that SCHC had 4.85 greater odds of having metritis<sup>45</sup> and higher incidence of endometritis in MF cases.<sup>46</sup>

Hypocalcaemia reduces contraction of smooth and skeletal muscles<sup>33</sup> and thereby reduces the teat sphincter muscle and myometrium contraction<sup>47</sup> that may prevent efficient teat closure along with impaired immune function leading to milk leakage which invites environmental pathogens to enter the udder and entrance of bacteria causing mastitis.<sup>17,34,44</sup> In addition, hypocalcemic cows tend to spend more time lying down which could increase teat end exposure to environmental pathogens.<sup>17,41,48</sup> The milk fever affected cows have been reported to be eight times more likely to develop mastitis than normal cows.<sup>34</sup> The dairy animals experience some degree of immune-suppression during the transition period<sup>49-52</sup> which might be due to decreased polymorphonuclear leukocytes, glycogen stores, decreased blood calcium level and increased non-esterified fatty acids (NEFA) and BHBA.<sup>49</sup> Hypocalcaemia reduces the ability of immune cells to respond to stimuli, thus contributing to infections such as mastitis.<sup>53</sup>

The SCHC has also been reported to be associated with the reduction of ruminal and abomasal motility that may cause a reduction of feed intake and weight loss especially in early lactation<sup>54-58</sup> and well exacerbate NEB in cows that are already underfed.<sup>41</sup>

Both the clinical and SCHC have been reported to be associated with decreased fertility especially increased number of services per conception, increased calving to conception interval, estrus cyclicity and pregnancy rate in dairy animals.<sup>41,45,59-61</sup> Hypocalcaemia results in reduced fertility in dairy cows due to its effect on uterine muscle function, slower uterine involution<sup>41,59</sup> and reduced blood flow to the ovaries.<sup>62</sup>

An increased risk of culling has been reported for cows with hypocalcaemia.<sup>35,63</sup> The serum calcium levels of 1.8 mmol/L,  $\leq 2.2$  mmol/L and  $\leq 2.3$  mmol/L resulted 3, 2.4 and 5.3 times more likely to be culled in the first 60 days, 1<sup>st</sup> and 2<sup>nd</sup> weeks of lactation, respectively.<sup>35,50</sup> Even the average productive life of a cow can be reduced 3.4 years with hypocalcemia.<sup>25,64</sup> The losses from SCHC have been estimated to be several times more than clinical cases in a herd.<sup>65</sup>

### Cutoff point (hypocalcemia)

The cut-off points for blood calcium level as  $\leq 8$  mg/dl ( $\leq 2.0$  mmol/L) for SCHC<sup>17,18,26</sup> and  $\leq 6.0$  mg/dl for clinical hypocalcemia<sup>17,18</sup> have been reported. However, there are other studies that apply different concentrations of the normal reference values for serum calcium of cattle have been defined as 8.5 to 10 mg / dl,<sup>17</sup> 2.0 to 2.5 mmol/L,<sup>26</sup> above 8.8 mg / dl,<sup>36</sup> 8.59 mg / dl<sup>37</sup> and  $2.12 \pm 0.50$  mM<sup>66</sup> but different cutoff values for SCHC have also been used as  $\leq 2.0$  mmol/L,<sup>18,26</sup>  $\leq 2.14$  mmol/L<sup>37</sup> and 1.88 mmol/L (7.5 mg/dl).<sup>67</sup>

Several different blood calcium thresholds that have been reported in different problems including  $\leq 1.93$  mmol/L for ketosis,  $\leq 2.05$  mmol/L for retained placenta and metritis and  $\leq 2.10$  mmol/L for displaced abomasum.<sup>45</sup> Blood calcium threshold  $\leq 1.97$  mmol/L for 2<sup>nd</sup> parity animals at 2 day-in-milk (DIM) associated with the risk of metritis and/or abomasal displacement, whereas blood calcium levels of primiparous cows are not associated with lower milk production at any of the DIM assessed.<sup>39</sup> It has also been suggested that this cut off point should be raised to 8.5 mg / dl (2.1 mmol/L) because cows below this concentration have developed metritis or metabolic disorders.<sup>68</sup> The lowest serum calcium concentration typically occurs within the few days after calving<sup>37</sup> and returns to normal within 2 to 3 days.<sup>69,70</sup> The comparatively higher levels of serum calcium ( $12.97 \pm 6.67$  mg/dl) and phosphorus ( $7.26 \pm 0.41$  mg/dl) have been reported in non-lactating than lactating cows as calcium ( $10.05 \pm 2.51$  mg/dl) and phosphorus ( $6.52 \pm 0.30$  mg/dl) in an inland report.<sup>71</sup>

### **Prevalence of SCHC**

The overall prevalence of SCHC was recorded in 30.0% lactating three types of cross-bred dairy cows with significantly ( $p < 0.05$ ) highest prevalence in HF  $\times$  L (32.11%) in comparison to SH  $\times$  L (15.0%) and JS  $\times$  L (20.0%) cross-bred cows. It appears that the prevalence of SCHC (30.0%) is at least 10 times more prevalence than clinical MF (2.97%) in dairy cattle in Bangladesh.<sup>23</sup> However, the overall lower prevalence of 30.0% SCHC recorded in cross-bred lactating dairy cows in Bangladesh in comparison to 50% in US dairy herds,<sup>18</sup> 40 to 50% in multiparous Iranian dairy herds,<sup>72</sup> 40.34% in lactating cows in Brazil<sup>73</sup> and 47.6% within 48 hours of parturition in Germany<sup>74</sup> and 30 to 50% on the day of calving elsewhere.<sup>18,75,76</sup>

Hypocalcaemia (MF/PP) affects high milk producing dairy animals in their transition period. Generally, pure exotic and their cross-bred are highly susceptible to hypocalcaemia due to high amount of colostrum and milk production. However, certain breeds of cattle have been reported to be more susceptible to hypocalcemia, particularly Channel Island, Swedish Red and White, and Jersey cattle.<sup>77</sup> The intestinal receptors for  $1,25(\text{OH})_2\text{D}_3$  are lower in Jersey than aged-matched Holsteins. Lower numbers of receptors would result in a loss of target tissue sensitivity to  $1,25(\text{OH})_2\text{D}_3$ , resulting in increased susceptibility to hypocalcemia. The risk of developing MF is higher in Jerseys and also increases with increasing parity and higher levels of milk production and thus advocating for special considerations when dairy cows fit these criteria.<sup>78</sup> The high prevalence rate of SCHC in adult dairy cattle at transition period<sup>79</sup> and therefore, prevention of hypocalcemia in peri-parturient animals has significant importance in cow health, finance and welfare and accordingly the SCHC has become a major topic of recent research works in dairy animals.

### **Influence of age on prevalence of SCHC**

The influence of age on the prevalence SCHC showed highest prevalence of SCHC in cows more than five years of age (37.16%) in comparison to lower aged cows less than four years (16.67%) and four to five years (25.33%). Age has a profound effect on susceptibility of dairy cows to hypocalcaemia and the risk of hypocalcaemia increases by approximately 9% per lactation. Older cows are affected by hypocalcaemia more common and more severe than



young cows. Older and over-conditioned cows have been reported to be at greater risk for experiencing MF,<sup>26</sup> whereas primiparous cows are much less likely to have low blood calcium levels near calving.<sup>80</sup> In addition to increased milk production from the 3<sup>rd</sup> lactation resulting high calcium demand, ageing also results in a diminished ability to mobilize calcium from bone stores and a decline in the active transport of calcium in the intestine as well as impaired production of  $1,25(\text{OH})_2\text{D}_3$  due to decrease number of  $1,25(\text{OH})_2\text{D}_3$  receptors. The hypocalcemia at calving is age related and most marked in cows from 3<sup>rd</sup> to 7<sup>th</sup> parity and it is infrequent at the first parturition.

### **Influence of parity on prevalence of SCHC**

The influence of parity on the prevalence of SCHC showed significantly ( $p < 0.05$ ) higher prevalence of SCHC at 4<sup>th</sup> parity (46.67%) and significantly ( $p < 0.05$ ) lower at 1<sup>st</sup> parity (16.67%) in comparison to other parities in dairy cows. These observations support the earlier reports that the prevalence of SCHC increases as the parity increases.<sup>18,81</sup> Primiparous cows have a lower risk of suffering from SCHC while multiparous cows have an increased risk. Multiparous cows, especially after the third parturition have an increased risk of suffering from severe SCHC. Significantly ( $p < 0.05$ ) higher prevalence of MF has been reported in Jerseys cows beyond their 4<sup>th</sup> parity (24.85%) than 2<sup>nd</sup> (5.9%), 3<sup>rd</sup> (6.49%) and 4<sup>th</sup> (8.73%) parities. Similarly, Holstein cows had higher MF beyond 4<sup>th</sup> parity (8.29%) than 2<sup>nd</sup> (1.43%), 3<sup>rd</sup> (1.82%) and 4<sup>th</sup> (2.91%) parities.<sup>18,81</sup>

The age and parity-associated susceptibility might be related calcium homeostatic mechanisms. With increasing age and parity, the hemostasis process is impeded and results in moderate to severe hypocalcaemia. It has been assumed that the number of vitamin D receptors in intestines decline with increasing age.<sup>82</sup> In addition, as animals age increase, the number of receptors for PTH on target tissue decline.<sup>83</sup>

### **Influence on lactation stages on prevalence of SCHC**

The prevalence of SCHC was found significantly ( $p < 0.05$ ) higher (94.44%) at the 1<sup>st</sup> week of lactation in comparison to higher lactation stages. Blood calcium levels remarkably declines in dairy cows around calving, with the lowest levels occurring about 12 to 24 hours after calving.<sup>17,53</sup> Blood samples tested at this stage can reveal the extent of hypocalcaemia experienced by a dairy herd.<sup>84</sup> The SCHC increased with age and 41, 49%, 51%, 54% and 42% have been reported at 2<sup>nd</sup> to 6<sup>th</sup> lactation cows, respectively.<sup>18,72</sup>

### **Influence SCHC on milk yield**

An association between the prevalence of SCHC and increased of milk yield was recorded with a significantly ( $p < 0.05$ ) higher prevalence of SCHC in cows produced more than 15 liter milk / day (68.57%) than cows produced less than 5.0 liter /day (2.50%), 5 to 10 liter/ day (20.0%) and 11 to 15 liter/day (30.43%) of milk. These results are in conformity with the findings of cows with SCHC produced an average of 5.7 kg / day more milk during 2, 3 and 4 weeks compared with control cows.<sup>85</sup>

### **Influence of BCS on SCHC**

This study also recorded higher BCS ( $> 3.5$ ) had higher prevalence of SCHC than the lower BCS ( $3 - 3.25$ ). This finding supports that the higher BCS ( $\geq 3.75$  out of 5.0) at calving developed up to 4 times more hypocalcemia in dairy cows.<sup>86</sup> Higher BCS at calving have a higher calcium output in milk and also results in decreased feed intake during gestation period due to reduced appetite in critical period and around calving which predisposes them to hypocalcaemia.<sup>87</sup>

### **Relationship between macro-minerals and Hypocalcemia**

The average reference values of serum micro-minerals of lactating dairy cows (calcium  $2.12 \pm 0.50$  mmol/L, phosphorus  $1.81 \pm 0.48$  mmol/L, magnesium  $0.97 \pm 0.52$  mmol/L) have been reported.<sup>66</sup> The SCHC has been reported to be associated with hypophosphatemia and hypermagnesemia in association with all the investigated risk factors including breeds, age, parity, lactation stages, milk yield and BCS in lactating dairy cows. These results suggest that the magnesium and phosphorus play an important role in calcium homeostasis.<sup>26</sup>

Animals fed diets containing less phosphorus than necessary to meet physiologic needs suffer hypophosphatemia. The hypophosphatemia is usually complicated by concurrent hypocalcemia, hypomagnesemia and in some cases hypoglycemia. At the onset of lactation in the dairy cow, production of colostrum and milk draw large amounts of phosphorus out of extracellular phosphorus pools, depressing blood phosphorus concentrations. Within a day or two of calving it is typical to find blood phosphorus concentration between 3.2 and 4 mg/dl in healthy cows. However, cows that develop hypocalcemia have low blood phosphorus concentrations that are even further depressed. Plasma inorganic phosphorus concentrations in cows with hypocalcemia are often between 1 and 2 mg/dl. Plasma phosphorus concentrations usually increase rapidly following treatment of the hypocalcemic cow with intravenous calcium solutions. Restoring normo-calcemia decreases parathyroid hormone secretion, which reduces urinary and salivary loss of phosphorus and stimulates resumption of gastro-intestinal motility, which in turn allows absorption of dietary phosphorus and reabsorption of salivary phosphorus secretions. Protected hypophosphatemia in some cows appears to be an important factor in some nonresponsive hypocalcemic cases. Unlike typical cases of parturient paresis, plasma phosphorus levels in these cows remains low, despite successful treatment of the hypocalcemia. Instituting a program to control hypocalcemia and MF generally is an effective means of preventing the low phosphorus downer cow syndrome.<sup>17</sup> In dry cows, high dietary levels of phosphorus ( $> 0.5\%$  DM intake) increase the serum level of inorganic phosphorus (IP) which has inhibitory effect on the renal enzyme ( $1\alpha$ -hydroxylase) that catalyzes the conversion of vitamin D into its active form ( $1,25(\text{OH})_2\text{D}_3$ ) and thereby predisposes cows to hypocalcaemia.<sup>88-90</sup> High dietary phosphorus has also been reported to have a negative effect on intestinal magnesium absorption which further makes periparturient cows susceptible to hypocalcaemia.<sup>91,92</sup>

Hypomagnesemia affects calcium metabolism by reducing PTH secretion in response to hypocalcaemia and by reducing ability of PTH stimulated cells to produce cyclic AMP resulting in failure to activate the target tissues to PTH.<sup>17,91</sup> Low magnesium levels in the diet

reduced calcium absorption in the gut. On the contrary, serum calcium and magnesium concentrations are negatively associated. Cows suffering from hypocalcaemia have higher serum magnesium level.<sup>74</sup> In a period of low serum calcium level, PTH is secreted into the blood. PTH secretion raises the threshold for renal magnesium excretion resulting in a higher serum magnesium concentration.<sup>17,93</sup> Hypocalcemia associated with hypermagnesemia may be due to in part to the suppressive effects of hypermagnesemia on PTH.<sup>94</sup>

### **Hypoglycemia and SCK**

Ketosis is a major metabolic disorder of dairy cows in early lactation which develops when dairy cows fall into a condition of excessive NEB caused by insufficient dietary intake and generous lactation and characterized by relatively high concentrations of the ketone bodies (acetoacetate, BHBA and acetone) and a concurrent low concentration of blood glucose.<sup>95</sup> The decreased DMI at pre-partum causes NEB and increases NEFA and BHBA concentrations that cause ketosis at early lactation.<sup>28</sup>

Bovine ketosis typically occurs in early lactation in both the clinical and sub-clinical forms. The CK is characterized by diminished appetite, decreased milk production, weight loss, hypoglycemia and hyperketonemia whereas SCK remain undetected clinically but have effects on productivity like clinical ketosis.<sup>96</sup> The SCK (more correctly called hyperketonemia) may be defined as increased concentrations of circulating ketone bodies without the presence of clinical signs of ketosis in lactating cows.<sup>97,98</sup> However, it can appear during the transition period, dry period or at calving or in early lactation where the highest incidence of SCK occurs within the first 2 to 3 weeks of lactation.<sup>99</sup>

The serum BHBA 1.2 to 1.4 mmol/L during the first and second weeks of calving have been associated with 1.5 to 2.4 kg milk loss daily.<sup>36,99-102</sup> The serum BHBA concentrations  $\geq 1.2$  mmol/L during the first week of calving have been reported to be associated with displacement of abomasum and metritis<sup>99,103</sup> and  $> 1.0$  mmol/L associated with significantly less conception rate after first AI.<sup>104</sup> The increased ketone bodies in cows immediately after parturition has a negative impact on health of cows and associated with reduced milk production.<sup>97,105</sup> The total cost of SCK per case per year resulted for 36% from a prolonged calving interval, 24% from reduced milk production, 19% from treatment, 14% from discarded milk and 6% from removal.<sup>27</sup>

### **Diagnosis and Cutoff value for ketosis**

Two major changes occur in the blood of ketosis affected dairy animals: (a) Hypoglycemia and (b) ketonemia. Ketosis occurs in early lactation because of the decrease in blood glucose levels, which leads to a high degree of fatty acid mobilization in the form for NEFA. The NEFAs are then oxidized by the liver, leading to ketone body (acetone, acetoacetate & BHBA) production. Diagnosis of ketosis is preferred by measuring acetoacetate and BHBA levels in the blood, urine or milk and blood glucose levels.

The average reference values of serum glucose ( $3.15 \pm 0.67$  mmol/L) with high milk yielding ( $3.01 \pm 0.65$  mmol/L), low milk yielding ( $3.18 \pm 0.69$  mmol/L) and dry cows ( $3.25 \pm 0.64$

mmol/L) in dairy cows have reported.<sup>66</sup> Significantly ( $p < 0.01$ ) lower plasma concentrations of glucose ( $4.23 \pm 0.58$  &  $2.74 \pm 0.51$  mmol/L) and calcium ( $2.18 \pm 0.14$  &  $2.08 \pm 0.15$  mmol/L) have been reported in ketosis affected lactating cows in comparison to healthy cows.<sup>106</sup>

Hypoglycemia, hyperketonemia, ketonuria and ketolactia are the biochemical characteristics of ketosis, and blood glucose levels are reduced from the normal of 50 mg/dl to 20 to 40 mg/dl.<sup>95</sup> The blood glucose level has been reported to be consistently low in cows at the time of their first ketosis diagnosis, even if the cows in very early lactation.<sup>107</sup> The CK cases are usually associated with plasma glucose concentrations less than 35 mg/dl and NEFA concentrations more than 1000  $\mu$ Eq/L.<sup>108</sup>

Blood glucose levels of  $44.8 \pm 2.2$  mg/dl in CK affected cows has been reported to be lower than SCK ( $51.6 \pm 2.3$ ) affected cows and blood glucose levels of both types of ketosis had lower than healthy ( $68.3 \pm 1.8$ ) cows.<sup>109</sup> However, blood glucose level  $\leq 44.0$  mg/dl has been considered as SCK in dairy cows.<sup>110</sup>

The 'gold standard' test for ketosis is serum BHBA because it is more stable in blood than acetone or acetoacetate<sup>80</sup> and its threshold levels for SCK ranges from 1.0 to 1.4 (1.2) mmol/L<sup>112-114</sup> whereas CK (reduced milk yield, lethargy and loss of appetite)  $> 3.0$  mmol/L.<sup>28,115-118</sup> However, some cows have been reported to be exposed with high levels of BHBA ( $> 3$  mmol/L) without showing any clinical signs and some other cows develop CK at reduced BHBA ( $< 3$  mmol/L) levels.<sup>13,98,119</sup>

The cow-level prevalence of hyperketonemia, hypoglycemia, and simultaneous hypoglycemia and hyperketonemia has been reported to be 20.0%, 13.8% and 6.2%, respectively. The herd level average prevalence of hypoglycemia within the subset of hyperketonemic cows only has reported as 30.6%.<sup>120</sup>

Comparative evaluation of three cow side tests for detection of SCK in early lactating cows showed that both the Ketostix and Keto-Test strips have provided acceptable results for screening individual cows on commercial dairies to detect SCK.<sup>121</sup> Urine dipstick test showed sensitivity and specificity of 86% and 100% respectively.<sup>122</sup> Using trace as a positive result for the Ketostix and blood BHBA  $\geq 1.2$  mmol/L as the gold standard for diagnosis of SCK resulted in sensitivity of 88% and specificity of 95%.<sup>123</sup> Non-availability of electronic cow side test (Precision Xtra), the ketone strips (Atena Medical Instrument Co., China) are a dipstick containing the salt nitroprusside which becomes pink in the presence of acetoacetate (AcAc) has been used in this study.

### **Overall prevalence of SCK**

The overall 25.0% prevalence of SCK was recorded in this study in lactating dairy cows based on the hypoglycemia and urinary ketone bodies tests. It appears that the prevalence of SCK (25.0%) is at least six times higher than the prevalence of clinical ketosis (3.75%) in dairy cattle in Bangladesh.<sup>23</sup> Insignificantly ( $p > 0.05$ ) higher prevalence of SCK was recorded in HF  $\times$  L (27.37%) than SH  $\times$  L (10.0%) and JS  $\times$  L (10.0%) cross-bred lactating dairy cows. These findings are in conformity with earlier report of an overall 25% prevalence of SCK detected by Rothera's test of which comparatively higher percentage of SCK was recorded in HF  $\times$  L (25.9%) than SH  $\times$  L (21.43%) cross-bred lactating cows.<sup>24</sup> However, reports on both the higher and lower

prevalence of SCK have been reported in lactating dairy cows elsewhere. The prevalence of SCK has been reported to be 7.5 to 14% in Canada and France,<sup>124,125</sup> 13.9% in Iran,<sup>107</sup> 18.0% in Turkey,<sup>109</sup> 36.6% in Europe,<sup>116</sup> 34% in Denmark,<sup>126</sup> and 29.3% in Hungarian dairy cattle.<sup>127</sup>

Prevalence of SCK ranges from 10 to 40% within 3 weeks of first lactation in small herds,<sup>80,99,100,101,114,116-118</sup> 7 to 14% in the first 60 days of lactation<sup>128</sup> but it varies from 8.3 to 40.1% with an average of 24.1% in large population of 8,902 dairy cows kept at 541 dairy farms of different countries in the world at the second week of lactation (> 95%) in dairy cows.<sup>129</sup> The prevalence of SCK has also been reported as range from 9 to 43% in the first 2 months of lactation<sup>100,101,112</sup> with the highest risk occurring within the first 21 days of lactation.<sup>80,99</sup> The average prevalence of SCK from 3 to 16 DIM has been reported to be 43% with range from 26 to 56%<sup>130</sup> with peak at 5 DIM.<sup>101</sup> However, it has also been reported in wide range between 7 and 73% in dairy herds.<sup>112,116,131,132</sup> Approximately 40% of all cows have been reported to had SCK at least once and > 90% had SCK in the first and second months after calving.<sup>133</sup> In comparison to the prevalence clinical ketosis ranges from 2 to 15% higher prevalence of 10-60% SCK have been reported.<sup>134</sup>

The higher prevalence of ketosis is significantly associated with exotic pure/cross-bred than native/non-descriptive cattle.<sup>95</sup> The higher prevalence of ketosis in exotic pure animals of high genetic potential because these animals are unable to withstand the pressures arising from the high nutritional demands generated by the production of high milk yield, which ultimately results in development of hypoglycemia that can remain as a challenge to the successful dairy farming business, eventually ruining the life of dairy farmers.<sup>95</sup>

### **Influence of Age on the prevalence of SCK**

Significantly ( $p < 0.01$ ) higher prevalence of SCK was recorded in more than 5 years of age (46.78%) in comparison to 4 to 5 years (4.0%) and < 4 years (2.78%) of aged lactating cows. These findings can be compared with the higher (29.31%) prevalence of SCK which has been reported in 8 to 9 years of age and lower (9.52%) in more than 9 years of age group of cows.<sup>135</sup> However, the prevalence of SCK is more important based on parity than age.

### **Influence of parity on the prevalence of SCK**

The significantly ( $p < 0.05$ ) highest prevalence of SCK was recorded at 4<sup>th</sup> parity (53.33%) and significantly ( $p < 0.01$ ) lower prevalence at the 1<sup>st</sup> (2.78%) and 2<sup>nd</sup> (4.00%) parities. A significant positive relationship between the parity and the probability of occurrence of ketosis has been reported. The probability of ketosis and positive cases has been reported to be peaked at the third and 4<sup>th</sup> lactations.<sup>127</sup> Therefore, hypoglycemia mainly occurred in multiparous cows with early-onset hyperketonemia whereas primiparous cows were at a lower risk for hypoglycemia.<sup>136</sup>

### **Influence of lactation stages on the prevalence of SCK**

The significantly ( $p < 0.01$ ) higher prevalence of SCK was recorded at 1<sup>st</sup> (77.78% and 2<sup>nd</sup> (56.67%) weeks of lactation stages in comparison to all recorded higher weeks of lactation stages (0 to 25.71%) in cross-bred lactating cows. The significantly higher prevalence of SCK



in the first two weeks of lactation recorded in this study supports the earlier reports.<sup>100,116,132,137-140</sup> However, comparatively higher rate of 90%<sup>133</sup> and lower rates of 47.2%<sup>141</sup> and 13.19%<sup>107</sup> SCK have been reported in lactating dairy cows elsewhere. The higher prevalence of ketosis has also been reported during 2<sup>nd</sup> (42.2%) and 3<sup>rd</sup> (24.8%) weeks post calving and accordingly it has been suggested that the 14 and 17 days after calving are the best time to detect CK and SCK in dairy cows.<sup>107</sup>

### **Influence of milk yield on the prevalence of SCK**

The prevalence of SCK was found higher with increased milk production which was found significantly ( $p < 0.01$ ) higher in cows produced milk more than 15 liter / day (57.14%) in comparison to < 5 liter (2.50%), 5 to 10 liter (13.33%) and 11 to 15 liter (26.09%) milk / day. These observations support the findings of average milk production for cows suffering from CK, SCK and healthy have been reported as 28, 35 and 45 kg per day, respectively.<sup>107</sup>

### **Influence of BCS on the prevalence of SCK**

The BCS prior to calving is an important risk factor for subsequent development of SCK during lactation.<sup>142,143</sup> Statistically ( $p > 0.05$ ) insignificant higher prevalence of SCK was recorded with > 3.5 BCS (30.0%) in comparison to BCS 3.0 - 3.25 (20.83%) in lactating cows. The higher BCS causes increased prevalence of SCK has been reported elsewhere.<sup>114,118,139,141,144</sup> The cows with BCS  $\geq 3.75$  out of 5.0 at calving have increased risk of ketosis and cows with excessive adipose stores (BCS  $\geq 3.75$  out of 5) at calving are at greater risk of developing CK.<sup>145</sup> A cow with a BCS  $\geq 3.5$  has been reported to be 2.5 times more likely to develop ketosis than cows with scores as lower than 3.25 at calving.<sup>144</sup> The change in BCS directly reflects the energy status of dairy cows<sup>146-148</sup> and has negative relationship between BCS to milk production in the middle lactation stage.<sup>149</sup>

### **Biochemical types of bovine ketosis**

The physiological tendency of dairy cows to respond to NEB by catabolism and utilization of their body reserves generates subclinical ketosis with biosynthesis of ketone bodies like acetone, acetoacetate and BHBA.<sup>99</sup> Blood glucose, insulin, non-esterified fatty acids (NEFA) and  $\beta$ -hydroxybutyrate (BHBA) are the most important biochemical constituents associated with the diagnosis of ketosis. Within four days post-partum in dairy cows, demands for glucose, amino acids and fatty acids due to milk production are two to five times higher than pre-partum requirements.<sup>30</sup> Higher demand of energy and nutrients for the synthesis of colostrum and milk combined together with decreased feed intake force the transition cows to undergo negative energy balance (NEB) and micro-nutrient deficiencies. NEB is a result of an imbalance between energy input and output because the energy requirements for milk production and maintenance exceed the available energy from feed intake.<sup>150,151</sup> The NEB (hypoglycemia) stimulates cows to mobilize body fat (an increase in lipolysis) in the form of NEFA.<sup>145,152</sup> The decrease of insulin and increase of NEFA showed the difficulty of dairy cows to cope with the energy demand during transition period.<sup>153</sup> The liver convert 15 to 20% of NEFA in ketone bodies (acetone, acetoacetate and BHBA), in triglycerols (TAGs) and

packaged into very low-density lipoproteins for transport back to the adipose tissue or stored as TAGs.<sup>153-155</sup> Excessive fat accumulation in the liver impairs normal liver function which may lead to hyperketonemia.<sup>128</sup>

It has been explained the theory of Type I and Type II ketosis based on differing in their onset and pathophysiology.<sup>128,156</sup> Type I ketosis was described as spontaneous or underfeeding ketosis, occurs 3 to 6 weeks post-partum when milk secretion is so extensive that the demand for glucose exceeds the capacity for gluconeogenesis in the liver. The plasma levels of glucose (hypoglycemia) and insulin are low and the levels of ketone bodies are high. Type II generally occurs earlier in lactation within the first two week of post-partum as a result of insulin resistance and excessive body fat mobilization prior to or at calving especially in obese cows and is also known as 'fat cow syndrome.' Blood insulin and glucose concentration are high whereas blood ketone concentrations are lower in Type II ketosis than in Type I.<sup>156</sup> Several studies evaluated the effects of HYK on milk yield<sup>36,99,1001,157,158</sup> and reproductive performance<sup>101,104,157</sup> within 2 weeks after parturition. The first 14 to 16 days after calving have been described as the main risk period for HYK.<sup>101,112</sup> But no consistent results have been found for the evaluation of associations between elevated post-partum BHBA and reproductive measures.<sup>159</sup> Moreover, other report suggested that early onset of HYK seems to be primary an adaptation response to a high metabolic load caused by higher milk production in the beginning of lactation without the negative side effects on health.<sup>136</sup>

The comparatively higher serum glucose level in lactating cows ( $63.02 \pm 6.67$  mg/dl) than the non-lactating cows ( $58.47 \pm 3.84$  mg/dl) has been reported from Bangladesh<sup>71</sup> which not only contradicts the higher serum glucose level reported in dry cows ( $69.8 \pm 3.7$  mg/dl) than lactating cows  $65.2 \pm 1.7$  mg/dl).<sup>160</sup> However, hyperglycemia in lactating cows could be explained probably due to stress, late pregnancy, insulin resistance Type II ketosis,<sup>136</sup> methodological error and other reasons which need to be investigated for further explanation.

### **Interaction of macro-minerals and glucose in metabolic diseases**

Blood macro-minerals and glucose levels have very important roles to health, growth, production and reproduction, immune and endocrine system functions. Homeostasis of calcium, phosphorus and magnesium is primarily affected by the very same homeostatic mechanisms and as a result, the changes in their concentrations are in most cases mutually linked.<sup>161</sup> The MF of dairy herds is biochemically characterized by hypocalcemia, frequently accompanied by hypophosphatemia and/or hypoglycemia but magnesium concentration may be normal, reduced or increased. However, in the course of disease the concentration of calcium and phosphorus decreases while the magnesium concentration increases which might be due to imbalance of ratios of these minerals.<sup>161</sup> The decrease in calcium level occurring at calving may results in the increase in magnesium<sup>162</sup> which may be due to increased calcium renal threshold, when in a response to reduced blood calcium level the PTH is excreted resulting in an increase in the magnesium renal threshold.<sup>17,163</sup>

The concurrent prevalence of SCHC and SCK (20.91%) was found significantly ( $p < 0.001$ ) higher in comparison to single occurrence of SCHC (8.64%) and SCK (5.05%) in lactating cross-bred cows. These results are in supports of findings that the hypocalcemic cows had



significantly higher association with ketosis on day 7 and day 35 post-partum.<sup>18,47,164</sup> Hypocalcaemia has been attributed to the occurrence of ketosis in dairy cows<sup>34</sup> and it has also reported that the hypocalcemic cows had 5.5 greater odds of having ketosis than normocalcemic cows.<sup>45</sup> It appears that the hypocalcaemia has impact on feed intake (reduction of appetite) and resulting NEB that might be the factor in promoting ketosis. Hypocalcaemia had elevated concentrations of NEFA and BHBA as indicators of increased lipid metabolism.<sup>18,37,68</sup> Hypocalcaemia may also deplete adipocyte calcium stores resulting in increased lipolysis.

### **Therapeutic trials against SCHC and SCK**

Treatment of clinical and subclinical MF and SCK are the main aimed at restoring calcium and glucose levels in the blood, respectively. Administration of intravenous glucose and/or propylene glycol orally and calcium salt orally is commonly used for restoring blood glucose and calcium levels in SCK and SCHC affected lactating cows. The blood calcium and glucose levels of both the SCHC and SCK affected cattle treated with the oral calcium salt and propylene glycol increased significantly ( $p < 0.001$ ) at 7 days of post-treatment in comparison to pre-treatment values, respectively. These findings are conformity with earlier reports.<sup>130,165,166</sup> Treatment with four doses of an oral calcium supplement (providing 50 g calcium/dose, given before calving, at calving, 12 hours post-calving and 24-hours post-calving) reduces the risk of clinical and sub-clinical Milk fever in dairy cows by about half.<sup>167,168</sup> However, oral supplementation with calcium chloride and calcium sulfate has been shown to have significant effects on improving calcium status in the period following calving.<sup>165,169</sup> In addition, the administration of the oral calcium bolus to high-producing cows leads to higher milk production.<sup>170</sup>

Cows treated with oral propylene glycol @ 300 ml / cow daily produced 0.23 kg more milk per milking in the first 30 days of lactation, for a total difference of 0.69 kg / cow per day. Propylene glycol treated cows have reported to be 1.5 times more likely to resolve their SCK and 0.54 times less likely to develop clinical ketosis. In addition, oral propylene glycol improves milk yield during early lactation in cows diagnosed with SCK.<sup>130</sup> It is identified that propylene glycol most likely reduces fatty acid mobilization from adipose tissue and by this mechanism can be protective against ketosis and fatty liver syndrome.<sup>171</sup>

### **CONCLUSIONS**

The sub-clinical metabolic disorders are the global issue and their prevalence can be varied in different herds and countries based on breeds, management and other risk factors. This study has recorded some reliable information on the prevalence of SCHC and SCK in lactating dairy crossbred cows in Bangladesh. These findings may be useful to set base-line data on levels of macro-minerals and glucose in dairy cows for different crossbred, ages, parity, BCS and lactation stages. The prevalence of SCHC was found 10 times and SCK six times greater than their clinical forms in lactating dairy cows. The high prevalence of SCHC and SCK should be viewed as a potential health risk to the transition cows. Culling, health problems, complications of parturition, loss of milk production, productive and reproductive problems are the common outcomes of these disorders. Measuring the calcium and glucose status of the fresh cow is the

first step in making intervention or management decisions in order to decrease the long-term consequences of SCHC and SCK in dairy cattle herds. Recently, an electronic BHBA handheld meter (Precision Xtra) has been validated for determination of BHBA from whole blood in dairy cows could be used in dairy herds in Bangladesh. Prevention of SCHC and SCK based on efficient diet, periodic screening for calcium, glucose and ketone bodies detection in different body fluids and BCS evaluation in dairy cattle are the best methods to early detection of these sub-clinical metabolic disorders in transition animals.

### ACKNOWLEDGEMENTS

The authors thank all the selected dairy farm owners and their staff members for use of their cows and facilities. The research work was supported by the National Science and Technology (NST) fellowship 2016-2017, The Ministry of Science and Technology, Bangladesh for the MS degree to the first author of this article. The authors gratefully acknowledge the authority and staff members for the BAU Central Laboratory for their cooperation during estimation of blood biochemical constituents of lactating dairy cows.

### CONFLICTING INTEREST

The authors declare that there is no conflict of interests regarding the publication of this article.

### REFERENCES

01. Datta AK, Haider MZ and Ghosh SK (2018). Economic analysis of dairy farming in Bangladesh. *Tropical Animal Health and Production* 51: 55-64 [doi: 10.1007/s11250-018-1659-7]
02. DLS (2019). Livestock economy at a glance, DLS. [dls.portal.gov.bd/sites/default/files/dls.portal.gov.bd/page/ee5f4621\\_fa3a\\_40ac\\_8bd9\\_898fb8ee4700/Livestock%20Economy%20at%20at%20glance%20%20%282017-2018%29.pdf](https://dls.portal.gov.bd/sites/default/files/dls.portal.gov.bd/page/ee5f4621_fa3a_40ac_8bd9_898fb8ee4700/Livestock%20Economy%20at%20at%20glance%20%20%282017-2018%29.pdf)
03. Kamal M.M. (2010). A review on cattle reproduction in Bangladesh. *International Journal of Dairy Science* 5: 245-252 [doi: 10.3923/ijds.2010.245.252]
04. Banglapedia (2015). Livestock. National Encyclopedia of Bangladesh. [en.banglapedia.org/index.php?title=Livestock](http://en.banglapedia.org/index.php?title=Livestock)
05. Parvez S (2018). Dairy industry in Bangladesh: Prospects and roadblocks. [thedailystar.net/supplements/dairy-industry-bangladesh-prospects-and-roadblocks-1584739](http://thedailystar.net/supplements/dairy-industry-bangladesh-prospects-and-roadblocks-1584739)
06. Anon. (2019). The role of dairy farm in the livestock sector of Bangladesh. <https://www.assignmentpoint.com/business/management/the-role-of-dairy-farm-in-the-livestock-sector-of-bangladesh.html>
07. Nath SK, Bhowmik DK, Rokonuzzaman M, Afrin K, Dash AK, and Alam MR (2016). Production performance of different cross breeds of milch cow in Mithapukur Upazila, Rangpur, Bangladesh. *International Journal of Advanced Multidisciplinary Research* 3: 29-33 [SOI: <http://s-o-org/1.15/ijarm-2016-3-6-5>]

08. Hamid MA, Rahman A, Zaman MA and Hossain KM (2017). Cattle genetic resources and their conservation in Bangladesh. *Asian Journal of Animal Sciences* 11: 54-64 [doi: 10.3923/ajas.2017.54.64]
09. Miazi OF, Hossain ME and Hassan MM (2007). Productive and reproductive performance of crossbred and indigenous dairy cows under rural conditions in Comilla, Bangladesh. *University Journal of Zoology Rajshahi University* 26: 67-70 [doi: 10.3329/ujzru.v26i0.702]
10. Naher L, Samad MA, Siddiki SHMF and Islam MT (2019). Relationship between blood metabolic profiles and milk yield associated with parity and stage of lactation in crossbred dairy cows in Bangladesh. *Journal of Veterinary Medical and One Health Research* 1: 185-199 [doi: 10.36111/jvmohr.2019.1(2).0011]
11. Haque SAMA (2007). Bangladesh: social gains from dairy development. [fao.org/3/i0588e/10588E03.htm](http://fao.org/3/i0588e/10588E03.htm)
12. Perween S, Singh A, Gupta M and Sahoo JK (2018). An update on milk fever and its economic consequences. *International Journal of Current Microbiology and Applied Sciences* 7: 2735-2742 [doi: 10.20546/ijcmas.2018.710.318]
13. Bruckmaier RM and Gross JJ (2017). Lactational challenges in transition dairy cows. *Animal Production Science* 57: 1471 [doi: 10.1071/AN16657]
14. Gross J, van Dorland HA, Bruckmaier RM and Schwarz FJ (2011). Performance and metabolic profile of dairy cows during a lactational and deliberately induced negative energy balance by feed restriction with subsequent realimentation. *Journal of Dairy Science* 94: 1820-1830 [doi: 10.3168/jds.2010-3707]
15. Bell AW (1995). Regulation of organic nutrient metabolism during transition from late pregnancy to early lactation. *Journal of Animal Science* 73: 2804-2819 [doi: 10.2527/1995.7392804x]
16. Butler WR and Smith RD (1989). Interrelationships between energy balance and post-partum reproductive function in dairy cattle. *Journal of Dairy Science* 72: 767-783 [doi: 10.3168/jds.S0022-0302(89)79169-4]
17. Goff JP (2008). The monitoring, prevention and treatment of milk fever and subclinical hypocalcaemia in dairy cows. *Veterinary Journal* 176: 50-57 [10.1016/j.tvjl.2007.12.020]
18. Reinhardt TA, Lippolis JD, McCluskey BJ, Goff JP and Horst RL (2011). Prevalence of subclinical hypocalcemia in dairy herds. *Veterinary Journal* 188: 122-124 [doi: 10.1016/j.tvjl.2010.03.025]
19. Mulligan FJ and Doherty ML (2008). Production diseases of the transition cow. *Veterinary Journal* 176: 3-9 [doi: 10.1016/j.tvjl.2007.12.018]
20. Drackley JK (1999). Biology of dairy cows during the transition period: The final frontier? *Journal of Dairy Science* 82: 2259-2273 [doi: 10.3168/jds.S0022-0302(99)75474-3]
21. Grummer RR (1995). Impact of changes in organic nutrient metabolism on feeding the transition dairy cow. *Journal of Animal Science* 73: 2820-2833 [doi: 10.2527/1995.7392820x]

## Sub-clinical milk fever and ketosis in crossbred cows

22. Mallard BA, Dekkers JC, Ireland MJ et al. (1998). Alteration in immune responsiveness during the peripartum period and its ramification on dairy cow and calf health. *Journal of Dairy Science* 81: 585-595 [doi: 10.3168/jds.S0022-0302(98)75612-7]
23. Samad MA (2019). A 50-year review on the prevalence of clinical diseases and disorders of cattle in Bangladesh. *Journal of Veterinary Medical and One Health Research* 1: 1-16 [doi:10.36111/jvmohr.2019.1(1). 0001]
24. Hossain SMS and Samad MA (2019). Prevalence of sub-clinical ketosis and its associated cow level risk factors in lactating dairy cross-bred cows in Bangladesh. *Journal of Veterinary Medical and One Health Research* 1 : 29-38 [doi: 10.36111/jvmohr.2019.1(1).0003]
25. Horst RL, Goff JP and Reinhardt TA (1997). Calcium and vitamin D metabolism during lactation. *Journal of Mammary Gland Biology and Neoplasia* 2: 253-263 [doi: 10.1023/A:1026384421273]
26. DeGaris PJ and Lean IJ (2008). Milk fever in dairy cows: A review of patho-physiology and control principles. *Veterinary Journal* 176: 58-69 [doi: 10.1016/j.tvjl.2007.12.029]
27. Mostert PF, Bokkers EAM, van Middelaar CE and Hogeveen H (2018). Estimating the economic impact of subclinical ketosis in dairy cattle using a dynamic stochastic stimulation model. *Animal* 12: 145-154 [doi: 10.1017/S1751731117001306]
28. LeBlanc SJ (2010). Monitoring metabolic health of dairy cattle in the transition period. *Journal of Reproduction and Development* 56 (suppl): S29-S35 [doi: 10.1262/jrd.1056s29]
29. Grummer RR, Mashek DG and Hayirli A (2004). Dry matter intake and energy balance in the transition period. *Veterinary Clinics of North America Food Animal Practice* 20: 447-470 [doi: 10.1016/j.cvfa.2004.06.013]
30. Bell AW (1995). Regulation of organic nutrient metabolism during transition from late pregnancy to early lactation. *Journal of Animal Science* 73: 2804-2818 [doi: 10.2527/1995.7392804x]
31. Corbett RB (2018). Monitoring ketosis in lactating dairy cows. dairyherd.com/article/monitoring-ketosis-lactating-dairy-cows
32. LeBlanc SJ, Lissemore KD, Kelton Df, Duffield TF and Leslie KE (2006). Major advances in disease prevention in dairy cattle. *Journal of Dairy Science* 89: 1267-1279 [doi: 10.3168/jds.S0022-0302(06)72195-6]
33. Murray RD, Horsfield JE, McCormick WD, Williams HJ and Ward D (2008). Historical and current perspectives on the treatment, control and pathogenesis of milk fever on dairy cattle. *Veterinary Record* 163: 561-565 [doi: 10.1136/vr.163.19.561]
34. Curtis CR, Erb HN, Sniffen CJ, Smith RD, Powers PA, Smith MC, White ME, Hillman RB and Pearson EJ (1983). Association of parturient hypocalcemia with eight periparturient disorders in Holstein cows. *Journal of the American Veterinary Medical Association* 183; 559-561 [PMID: 6618988]

35. Seifi HA, LeBlanc SJ, Leslie KE and Duffield TF (2011). Metabolic predictors of post-partum disease and culling risk in dairy cattle. *Veterinary Journal* 188: 216-220 [doi: 10.1016/j.tvjl.2010.04.007]
36. Chapinal N, Carson ME, LeBlanc SJ, Leslie KE, Godden S, Capel M, Santos JEP, Overton MW and Duffield TF (2012). The association of serum metabolites in the transition period with milk production and early-lactation reproductive performance. *Journal of Dairy Science* 95: 1301-1309 [doi: 10.3168/jds.2011-4724]
37. Martinez N, Risco CA, Lima FS, Bisinotto RS, Greco LF, Ribeiro ES, Maunsell F, Galvao K and Santo JEP (2012). Evaluation of periparturient calcium status, energetic profile and neutrophil function in dairy cows at low or high risk of developing uterine disease. *Journal of Dairy Science* 95: 7158-7172 [doi: 10.3168/jds.2012-5812]
38. Kocabagli N (2018). Prevention of milk fever: A herd health approach to dairy cow nutrition. *Archives of Animal Husbandry and Dairy Science* 1: 2018.AAHDS.MS.ID.000502 [doi: 33552/AAHDS.2018.01.000502]
39. Neves RC, Leno BM, Bach KD and McArt JAA (2018). Epidemiology of subclinical hypocalcemia in early-lactation Holstein dairy cows: The temporal associations of plasma calcium concentration in the first 4 days in milk with disease and milk production. *Journal of Dairy Science* 101: 9321-9331 [doi: 10.3168/jds.2018-14587]
40. Madison JB and Troutt HF (1988). Effects of hypocalcemia on abomasal motility. *Research in Veterinary Science* 44: 264-266 [PMID:3387682]
41. Mulligan FJ, Grady L, Rice DA and Doherty ML (2006). Production diseases of the transition cow: Milk fever and subclinical hypocalcaemia. *Irish Veterinary Journal* 59: 697-702
42. Erb HN, Smith RD, Oltenacu PA, Guard CL, Hillman RB, Powers PA, Smith MC and White ME (1985). Path model of reproductive disorders and performance, milk fever, mastitis, milk yield and culling in Holstein cows. *Journal of Dairy Science* 68: 3337-3349 [doi: 10.3168/jds.S0022-0302(85)81244-3]
43. Correa MT, Erb H and Scarlett J (1993). Path analysis for seven post-partum disorders of Holstein cows. *Journal of Dairy Science* 76: 1305-1312 [doi: 10.3168/jds.S0022-0302(93)77461-5]
44. Goff JP and Horst RL (1997). Effects of the addition of potassium or sodium but not calcium to pre-partum ratios on milk fever in dairy cows. *Journal of Dairy Science* 80: 176-186 [doi: 10.3168/jds.s0022-0302(97)75925-3]
45. Rodriguez EM, Aris A and Bach A (2017). Associations between subclinical hypocalcemia and post-parturient diseases in dairy cows. *Journal of Dairy Science* 100: 7427-7434 [doi: 10.3168/jds.2016-12210]
46. Whiteford LC and Sheldon IM (2005). Association between clinical hypocalcemia and post-partum endometritis. *Veterinary Record* 157: 202-203 [doi: 10.1136/vr.157.7.202]

## Sub-clinical milk fever and ketosis in crossbred cows

47. Chamberlin WG, Middleton JR, Spain JN, Johnson GC, Ellersieck MR and Pithua P (2013). Subclinical hypocalcemia, plasma biochemical parameters, lipid metabolism, post-partum disease and fertility in post-parturient dairy cows. *Journal of Dairy Science* 96: 7001-7013 [doi: 10.3168/jds.2013-6901]
48. Goff JP (2003). Managing the transition cow- considerations for optimizing energy and protein balance and immune function. *Cattle Practice* 11: 52-63
49. Galvao KN (2012). Association between immune function and development of uterine disease in dairy cows. *Animal Reproduction* 9: 318-322
50. Duffield T (2005). Impact of sub-clinical metabolic disease on risk of early lactation culling. *Journal of Dairy Science* 88 (Suppl. 1): 199-200
51. Wyle FA and Kent JR (1977). Immunosuppression by sex steroid hormones. The effect upon PHA- and PPD-stimulated lymphocytes. *Clinical and Experimental Immunology* 27: 407-415 [PMID: 862230]
52. Franklin ST, Young JW and Nonnecke BJ (1991). Effects of ketones, acetate, butyrate and glucose on bovine lymphocyte proliferation. *Journal of Dairy Science* 74: 2507-2514 [doi: 10.3168/jds.S0022-0302(91)78428-2]
53. Kimura K, Reinhardt TA and Goff JP (2006). Parturition and hypocalcemia blunt calcium signals in immune cells of dairy cattle. *Journal of Dairy Science* 89: 2588-2595 [doi: 10.3168/jds.S0022-0302(06)72335-9]
54. Mulligan FJ, O'Grady L, Rice DA and Doherty ML (2006). A herd health approach to dairy cow nutrition and production diseases of the transition cow. *Animal Reproduction Science* 96: 331-353 [doi: 10.1016/j.anireprosci.2006.08.011]
55. Hansen SS, Norgaard P, Pedersen C, Jorgensen RJ, Mellau LSB and Enemark JD (2003). The effect of sub-clinical hypocalcemia induced by Na<sub>2</sub>EDTA on the feed intake and chewing activity of dairy cows. *Veterinary Research Communications* 27: 193-205 [doi: 10.1023/a:1023340506782]
56. Daniel RC (1983). Motility of the rumen and abomasum during hypocalcemia. *Canadian Journal of Comparative Medicine* 47: 276-280 [PMID: 6416656]
57. Jorgensen RJ, Nyengaard NR, Hara S, Enemark JM and Andersen PH (1998). Rumen motility during induced hyper-and hypocalcemia. *Acta Veterinaria Scandinavica* 39: 331-338 [PMID: 9787496]
58. Caixeta LS, Ospina PA, Capel MB and Nydam DV (2015). The association of sub-clinical hypocalcemia, negative energy balance and disease with bodyweight change during the first 30 days post-partum in dairy cows milked with automatic milking systems. *Veterinary Journal* 204: 150-156 [doi: 10.1016/j.tvjl.2015.01.021]
59. Borsberry S and Dobson H (1989). Peri-parturient diseases and their effect on reproductive performance in five dairy herds. *Veterinary Record* 124: 217-219 [doi: 10.1136/vr.124.9.217]

60. Ribeiro ES, Lima FS, Greco LF, Bisinotto RS, Monteiro APA, Favoreto M, Ayres H, Marsola RS, Martinez N, Thatcher WW and Santos JE (2013). Prevalence of peri-parturient diseases and effects on fertility of seasonally calving grazing dairy cows supplemented with concentrates. *Journal of Dairy Science* 96: 5682-5697 [doi: 10.3168/jds.2012-6335]
61. Caixeta LS, Ospina PA, Capel MB and Nydam DV (2017). Association between subclinical hypocalcemia in the first 3 days of lactation and reproductive performance of dairy cows. *Theriogenology* 94: 1-7 [doi: 10.1016/j.theriogenology.2017.01.039]
62. Jonsson NN and Daniel RC (1997). Effects of hypocalcemia on blood flow to the ovaries of the sheep. *Transboundary and Emerging Diseases* 44: 281-287 [doi: 10.1111/j.1439-0442.1997.tb01112.x]
63. Roberts T, Chapinal N, LeBlanc SJ, Kelton DF, Dubuc J and Duffield TF (2012). Metabolic parameters in transition cows as indicators for early lactation culling risk. *Journal of Dairy Science* 95: 3057-3063 [doi: 10.3168/jds.2011-4937]
64. Curtis CR, Erb MN, Sniffen CJ and Smith RD (1984). Epidemiology of parturient paresis: predisposing factors with emphasis on dry cow feeding and management. *Journal of Dairy Science* 67: 817 - 825[ doi: 10.3168/jds.S0022-0302(84)81372-7]
65. Houe H, Qstergaard S, Thilsing-Hansen T, Jorgensen RJ, Larsen T, Sorensen JT, Agger JF and Blom JY (2001). Milk fever and subclinical hypocalcaemia: an evaluation of parameters on incidence risk, diagnosis, risk factors and biological effects as input for a decision support system for disease control. *Acta Veterinaria Scandinavica* 42: 1-29
66. Galvao KN, Neto AV, Pena G, Bittar J and Ibarbia L (2019). Comparing the urine ketone strip test and the handheld ketone meter to diagnosis ketosis in early lactation dairy cows. [edis.ifas.ufl.edu/pdffiles/VM/VM18600.pdf](https://edis.ifas.ufl.edu/pdffiles/VM/VM18600.pdf)
67. Goff JP, Littledike ET and Horst RL (1986). Effect of synthetic bovine parathyroid hormone in dairy cows: Prevention of hypocalcaemic parturient paresis. *Journal of Dairy Science* 69: 2278-2289 [doi: 10.3168/jds.S0022-0302(86)80666-X]
68. Martinez N, Sinedino LD, Bisinotto RS, Ribeiro ES, Gomes GC, Lima FS, Greco LF, Risco CA, Galvao KN, Taylor-Rodriguez D and Driver JP (2014). Effect of induced subclinical hypocalcemia on physiological responses and neutrophil function in dairy cows. *Journal of Dairy Science* 97: 874-887 [doi: 10.3168/jds.2013-7408]
69. Ramberg CF, Johnson EK, Fargo RD and Kronfeld DS (1984). Calcium homeostasis in cows with special reference to parturient hypocalcemia. *American Journal of Physiology* 246: 698-704 [doi: 10.1152/ajpregu.1984.246.5.R698]
70. Horst RL, Goff JP and Reinhardt TA (1994). Calcium and vitamin D metabolism in the dairy cow. *Journal of Dairy Science* 77: 1936-1951 [doi: 10.3168/jds.S0022-0302(94)77140-X]



## Sub-clinical milk fever and ketosis in crossbred cows

71. Sarker MS, Ahaduzzaman M, Sayeed MA, Sarker R, Nanno MA, Mannan A and Hossain MB (2015). Comparison of some serum biochemical parameters between lactating and non-lactating dairy cows in selected dairy farms of Chittagong district of Bangladesh. *Asian Journal of Medical and Biological Research* 1: 259-264 [doi: 10.3329/ajmbr.v1i2.25620]
72. Seifi HA, Mohri M and Zadeh JK (2004). Use of pre-partum urine pH to predict the risk of milk fever in dairy cows. *Veterinary Journal* 167: 281-285 [doi: 10.1016/S1090-0233(03)00114-X]
73. Da Silva DC, Femandes BD, Dos SLJM, Rodrigues GP, Dias DLB, de Oliveira SEJ and Filho HAM (2019). Prevalence of subclinical hypocalcemia in dairy cows in the Sousa city micro-region, Paraiba state. *Tropical Animal Health and Production* 51: 221-227 [doi: 10.1007/s11250-018-1680-x]
74. Venjakob PL, Borchardt S and Heuwieser W (2017). Hypocalacemia- cow-level prevalence and preventive strategies in German dairy herds. *Journal of Dairy Science* 100: 9258-9266 [doi: 10.3168/jds.2016-12494]
75. Roche JR (2003). The incidence and control of hypocalcaemia in pasture-based systems. *Acta Veterinaria Scanddinavica* 97: 141-144
76. Perez EMR (2015). Study of cow subclinical hypocalcemia and development of new tools for its diagnostic and prevention. [ddd.uab.cat/pub/tesis/2015/hdl\\_10803\\_323908/emrp1de1.pdf](http://ddd.uab.cat/pub/tesis/2015/hdl_10803_323908/emrp1de1.pdf)
77. Kusumanti E, Agger JF, and Jensen K (1993). Association between incidence risk of milk fever and lactation number, breed and season. *Acta Veterinaria Scanddinavica* S89: 141
78. Chiwome B, Kandiwa E, Mushonga B, Sajeni S and Habarugira G (2017). A study on the incidence of milk fever in Jersey and Holstein cows at a dairy farm in beatrice, Zimbabwe. *Journal of South African Veterinary Association* 88: 1457 [doi: 10.4102/jsava.v88i0.1457]
79. Horst RL, Goff JP and Reinhardt TA (2003). Role of vitamin D in calcium homeostasis and its use in prevention of bovine peri-parturient paresis. *Acta Veterinaria Scanddinavica Suppl* 97: 35-55
80. Oetzel GR (2004). Monitoring and testing dairy herds for metabolic disease. *Veterinary Clinics North America Food Animal Practice* 20: 651-674 [doi: 1016/j.cvfa.2004.06.006]
81. Curtis CR, Erb HN, Sniffen CJ, Smith RD and Kronfeld DS (1985). Path analysis of dry period nutrition, postpartum metabolic and reproductive disorders and mastitis in Holstein cows. *Journal of Dairy Science* 68: 2347-2360 [doi: 10.3168/jds.S0022-0302(85)81109-7]
82. Horst RL, Goff JP and Reinhardt TA (1990). Advancing age results in reduction of intestinal and bone 1,25-dihydroxyvitamin D receptor. *Endocrinology* 126: 1053-1057 [doi: 10.1210/endo-126-2-1053]
83. Hanai H, Brennan DP, Cheng I, Goldman ME, Chorev M, Levine MA, Sacktor B and Liang CT (1990). Down regulation of parathyroid hormone receptors in renal membranes from aged rats. *American Journal of Physiology- Renal Physiology* 259: F444-450 [doi: 10.1152/ajprenal.1990.259.3.F444]

84. Goff JP (2014). Calcium and magnesium disorders. *Veterinary Clinics of North America: Food Animal Practice* 30: 359-381 [doi: <https://doi.org/10.1016/j.cvfa.2014.04.003>]
85. Jawor PE, Huzzey JM, LeBlanc SJ and von Keyserlingk MA (2012). Associations of subclinical hypocalcemia at calving with milk yield and feeding, drinking and standing behaviors around parturition in Holstein cows. *Journal of Dairy Science* 95: 1240-1248 [doi: 10.3168/jds.2011-4586]
86. Ostergaard S, Sorensen J and Houe H (2003). A stochastic model simulating milk fever in a dairy herd. *Preventive Veterinary Medicine* 58: 125-143 [doi: 10.1016/s0167-5877(03)00049-7]
87. Harris D, Rukkwamsuk T and Wensing T (1999). Relationship between over feeding and over conditioning in the dry period and the problems of high producing dairy cows during the post-parturient period. *Veterinary Quarterly* 21: 71-77 [doi: 10.1080/01652176.1999.9694997]
88. Jorgensen NA (1974). Combating milk fever (dairy cattle, metabolic disorders). *Journal of Dairy Science* 57: 933-944 [doi: 10.3168/jds.S0022-0302(74)84989-1]
89. Reinhardt TA and Conard HR (1980). Mode of action of pharmacological doses of cholecalciferol during parturient hypocalcemia in dairy cows. *Journal of Nutrition* 110: 1589-1596 [doi: 10.1093/jn/110.8.1589]
90. Grunberg W (2014). Treatment of phosphorus balance disorders. *Veterinary Clinics: Food Animal Practice* 30: 383-408 [doi: 10.1016/j.cvfa.2014.03.002]
91. Goff JP (2004). Macromineral disorders of the transition cow. *Veterinary Clinics: Food Animal Practice* 20: 471-494 [doi: 10.1016/j.cvfa.2004.06.003]
92. Schnewille JT, Klooster AT and Beynen AC (1994). High phosphorus intake depresses apparent magnesium absorption in pregnant heifers. *Journal of Animal Physiology and Animal Nutrition* 71: 15-21
93. Martin-Tereso J and Martens H (2014). Calcium and magnesium physiology and nutrition in relation to the prevention of milk fever and tetany (dietary management of macro-minerals in preventing disease). *Veterinary Clinics: Food Animal Practice* 30: 643-670 [doi: 10.1016/j.cvfa.2014.07.007]
94. Cholt IN, Steinberg SF, Tropper PJ, Fox HE, Segre GV and Bilezikian JP (1984). The influence of hypermagnesemia on serum calcium and parathyroid hormone levels in human subjects. *New England Journal of Medicine* 10: 1221-1225 [doi: 10.1056/NEJM198405103101904]
95. Madreseh-Ghahfarokhi S, Dehghani-Samani A and Dehghani-Samani A (2018). Ketosis (acetonaemia) in dairy cattle farms: practical guide based on importance, diagnosis, prevention and treatments. *Journal of Dairy, Veterinary and Animal Research* 7: 299-302 [doi: 10.15406/jdvar.2018.07.00230]
96. Baird GD (1982). Primary ketosis in the high-producing dairy cow: clinical and subclinical disorders, treatment, prevention and outlook. *Journal of Dairy Science* 65: 1-10 [doi: 10.3168/jds.s0022-0302(82)82146-2]

## Sub-clinical milk fever and ketosis in crossbred cows

97. Duffield T (2000). Subclinical ketosis in lactating dairy cattle. *Veterinary Clinics of North America: Food Animal Practice* 16: 231-253 [doi: 10.1016/S0749-0720(15)30103-1]
98. Andersson L (1988). Subclinical ketosis in dairy cows. *Veterinary Clinics of North America Food Animal Practice* 4: 233-251 [doi: 10.1016/S0749-0720(15)31046-X]
99. Duffield TF, Lissemore KD, McBride BW and Leslie KE (2009). Impact of hyperketonemia in early lactation dairy cows on health and production. *Journal of Dairy Science* 92: 571-580 [doi: 10.3168/jds.2008-1507]
100. Dohoo IR and Martin SW (1984). Subclinical ketosis: prevalence and association with production and disease. *Canadian Journal of Comparative Medicine* 48: 1-5
101. McArt JAA, Nydam DV and Oetzel GR (2012). Epidemiology of subclinical ketosis in early lactation dairy cattle. *Journal of Dairy Science* 95: 5056-5066 [doi: 10.3168/jds.2012-5443]
102. Gustafsson AH, Andersson L and Emanuelson U (1993). Effect of hyperketonemia, feeding frequency and intake of concentrate and energy on milk yield in dairy cows. *Animal Production* 56: 51-60 [doi: 10.1017/S0003356100006152]
103. Ospina PA, Nydam DV, Stokol T and Overton TR (2010). Evaluation of nonesterified fatty acids and beta-hydroxybutyrate in transition dairy cattle in the northeastern United States: critical thresholds for prediction of clinical diseases. *Journal of Dairy Science* 93: 546-554 [doi: 10.3168/jds.2009-2277]
104. Walsh RB, Walton JS, Kelton DF, LeBlanc SJ, Leslie KE and Duffield TF (2007). The effect of subclinical ketosis in early lactation on reproductive performance of post-partum dairy cows. *Journal of Dairy Science* 90: 2788-2796 [doi: 10.3168/jds.2006-560]
105. Raboisson D, Mounie M and Maigne E (2014). Diseases, reproductive performance, and changes in milk production associated with subclinical ketosis in dairy cows: A meta-analysis and review. *Journal of Dairy Science* 97: 7547-7563 [doi: 10.3168/jds.2014-8237]
106. Yang W, Zhang B, Xu C, Zhang H and Xia C (2019). Effect of ketosis in dairy cows on blood biochemical parameters, milk yield and composition and digestive capacity. *Journal of Veterinary Research* 63: [doi: 10.2478/jvetres-2019-0059]
107. Samiei A, Liang JB, Ghorbani GR, Hirooka H, Ansari-Mahyari S and Sadri H (2013). Prevalence of ketosis and its correlation with lactation stage, parity and peak of milk yield in Iran. *Asian Journal of Animal and Veterinary Advances* 8: 604-612 [doi: 10.3923/ajava.2013.604.612]
108. Herdt TH and Gerloff BJ (2008). Ketosis. Chapter 36. In: Anderson DE and Rings M (2008). *Current Veterinary Therapy- E-Book: Food Animal Practice*. Elsevier Health Sciences, Saunders Elsevier, St. Louis, Missouri 63146, USA pp. 135-137
109. Senturk S, Cihan H, Mecitoglu Z, Catik S, Akgul GD, Kasap S and Topal O (2016). Prevalence of ketosis in dairy herds in Marmara, Aegean and Mediterranean regions of Turkey. *Ankara University Veteriner Fakulesi Dergisi* 63: 283-288

110. Radostits OM, Gay CC, Hinchcliff KW and Constable PD (2007). Metabolic disturbances. *Veterinary Medicine*. 10<sup>th</sup> edn., Baillire Tindal Publisher, London pp. 1627-1642
111. Tehrani-Sharif M, Hadadi M, Noughabi HH, Mohammadi A, Rostami F and Sharifi H (2012). Bovine subclinical ketosis in dairy herds in Nishaboor, Iran. *Comparative Clinical Pathology* 21: 1637-1641 [doi: <https://doi.org/10.1007/s00580-011-1340-2>]
112. Duffield TF, Sandals D, Leslie KE et al. (1998). Efficacy of monensin for the prevention of subclinical ketosis in lactating dairy cows. *Journal of Dairy Science* 81: 2866-2873 [doi: 10.3168/jds.S0022-0302(98)75846-1]
113. Iwersen M, Falkenberg U, Voigtsberger R, Forderung D and Heuwieser W (2009). Evaluation of an electronic cowside test to detect subclinical ketosis in dairy cows. *Journal of Dairy Science* 92: 2618-2624 [doi: 10.3168/jds.2008-1795]
114. Rollin E, Berghaus RD, Rapnicki P, Godden SM and Overton MW (2010). The effect of injectable butaphosphan and cyanocobalamin on post-partum serum beta-hydroxybutyrate, calcium and phosphorus concentrations in dairy cattle. *Journal of Dairy Science* 93: 978-987 [doi: 10.3168/jds.2009-2508]
115. LeBlanc SJ, Duffield TF, Leslie KE, Bateman KG, TenHag J, Walton JS and Johnson WH (2002). The effect of prepartum injection of vitamin E on health in transition dairy cows. *Journal of Dairy Science* 85: 1416-1426 [doi: 10.3168/jds.S0022-0302(02)74209-4]
116. Suthar VS, Canelas-Raposo J, Deniz A, and Heuwieser W (2013). Prevalence of subclinical ketosis and relationships with post-partum diseases in European dairy cows. *Journal of Dairy Science* 96: 2925-2938 [doi: 10.3168/jds.2012-6035]
117. Compton CW, McDougall S, Young L and Bryan MA (2014). Prevalence of subclinical ketosis in mainly pasture-grazed dairy cows in New Zealand in early lactation. *New Zealand Veterinary Journal* 62: 30-37 [doi: 10.1080/00480169.2013.823828]
118. Garro CJ, Mian L and Roldan MC (2014). Subclinical ketosis in dairy cows: prevalence and risk factors in grazing production system. *Journal of Animal Physiology and Animal Nutrition* 98: 838-844 (doi: 10.1111/jpn.12141]
119. Zbinden RS, Falk M, Munger A, Dohme-Meier F, van Dorland HA, Bruckmaier RM and Gross JJ (2017). Metabolic load in dairy cows kept in herbage-based feeding system and suitability of potential markers for compromised well-being. *Journal of Animal Physiology and Animal Nutrition* 101: 767-778 [doi: 10.1111/jpn.12498]
120. Dubuc J and Buczinski S (2018). Cow-and herd-level prevalence of hypoglycemia in hyperketonemic post-partum dairy cows. *Journal of Dairy Science* 101: 3374-3379 [doi: 10.3168/jds.2017-13773]
121. Carrier J, Stewart S, Godden S, Fetrow J and Rapnicki P (2004). Evaluation and use of three cowside tests for detection of subclinical ketosis in early post-partum cows. *Journal of Dairy Science* 87: 37-25-3735 [doi: 10.3169/jds.S0022-0302(04)73511-0]

## Sub-clinical milk fever and ketosis in crossbred cows

122. Ravi R, Vijayakumar G, Mohanambal K, Sivaraman S and Reddy BS (2019). Cow side tests for detection of ketosis in dairy cows. *Indian Veterinary Journal* 96: 71-72
123. Galvao KN, Neto AV, Pena G, Bittar J and Ibarbia L (2018). Comparing the urine ketone strip test and the handheld ketone meter to diagnose ketosis in early lactation dairy cows. Publication # VM 186. edis.ifas.ufl.edu/vm186
124. Geishauser T, Leslie K, Tenhag J and Bashiri A (2000). Evaluation of eight cow-side ketone tests in milk for detection of subclinical ketosis in dairy cows. *Journal of Dairy Science* 83: 296-299 [doi: 10.3168/jds.S0022-0302(00)74877-6]
125. Enjalbert F, Nicot MC, Bayourthe C and Moncoulon R (2001). Ketone bodies in milk and blood in dairy cows: Relationship between concentrations and utilization for detection of subclinical ketosis. *Journal of Dairy Science* 84: 583-589 [doi: 10.3168/jds.S0022-0302(01)74511-0]
126. Ingvarsen KL (2006). Feeding and management related diseases in the transition cow: physiological adaptations around calving and strategies to reduce feeding-related diseases. *Animal Feed Science and Technology* 126:175-213 [doi: 10.1016/j.anifeedsci.2005.08.003]
127. Hejel P, Zechner G, Csorba C and Konyves L (2018). Survey of ketolactia, determining the main predisposing management factors and consequences in Hungarian dairy herds by using a cow-side milk test. *Veterinary Record* 5(1):e000253 [doi: 10.1133/vetreco-2017-000253]
128. Herdt TH (2000). Ruminant adaptation to negative energy balance. Influences on the etiology of ketosis and fatty liver. *Veterinary Clinics of North America: Food Animal Practice* 16: 215-230 [doi: 10.1016/s0749-0720(15)30102-x]
129. Brunner N, Groeger S, Raposo JC, Bruckmaier RM and Gross JJ (2019). Prevalence of subclinical ketosis and production diseases in dairy cows in Central and South America, Africa, Asia, Australia, New Zealand and Eastern Europe. *Translational Animal Science* 3: 84-92 [doi: 10.1093/tas/txy102]
130. McArt JA, Nydam DV, Ospina PA et al. (2011). A field trial on the effect of propylene glycol on milk yield and resolution of ketosis in fresh cows diagnosed with subclinical ketosis. *Journal of Dairy Science* 94: 6011-6-20 [doi: 10.3168/jds.2011-4463]
131. McKay S (2012). Focus on subclinical ketosis at World Buiatrics Conference. *Large Animal Review* 18: 22-23
132. Duffield TF, Kelton DF and Leslie KE (1997). Use of test day milk fat and milk protein to detect sub-clinical ketosis in dairy cattle in Ontario. *Canadian Veterinary Journal* 38: 713-718
133. Geishauser T, Leslie K, Kelton D and Duffield T (2001). Monitoring sub-clinical ketosis in dairy herds. *Compendium: Continuing Education* 23: 65-71
134. Itle AJ, Huzzey JM, Weary DM and Von Keyserlingk MA (2015). Clinical ketosis and standing behavior in transition cows. *Journal of Dairy Science* 98: 128-134 [doi: 10.3168/jds.2014-7932]

135. Mohammed N, Jaiswal M and Bihani DK (2019). Prevalence of subclinical and clinical ketosis in cattle in and around Bikaner. *Journal of Biological Rhythm Research* [doi: 10.1080/09291016.2019.1629167]
136. Ruoff J, Borchardt S, Mahrt A and Heuwieser W (2016). Effects of hyperketonemia within the first six weeks of lactation on milk production and reproductive performance. *Journal of Advances in Dairy Research* 4 : 165 [doi: 10.4172/2329-888X.1000165]
137. Nielen M, Aarts MGA, Jonkers AGM, Wensing T and Schukken YH (1994). Evaluation of two cow-side tests for the detection of subclinical ketosis in dairy cows. *Canadian Veterinary Journal* 35: 229-232
138. Sakha M, Ameri M, Sharifi H and Taheri I (2007). Bovine sub-clinical ketosis in dairy herds in Iran. *Veterinary Research Communications* 31: 673-679 [doi: 10.1007/s11259-007-0053-0]
139. Erb HN and Grohn YT (1988). Epidemiology of metabolic disorders in the periparturient dairy cow. *Journal of Dairy Science* 71: 25-57 [doi: 10.3168/jds.S0022-0302(88)79845-8]
140. Sepulveda-Varas P, Weary DM, Noro M and von Keyserlingk MA (2015). Transition diseases in grazing dairy cows are related to serum cholesterol and other analytes. *PLoS One* 10: e0122317 [doi: 10.1371/journal.pone.0122317]
141. Vanholder T, Papen J, Bemers R, Vertenten G and Berge AC (2015). Risk factors for subclinical and clinical ketosis and association with production parameters in dairy cows in the Netherlands. *Journal of Dairy Science* 98: 880-888 [doi: 10.3168/jds.2014-8362]
142. Gearhart MA and Curtis CR (1990). Relationship of changes in condition score to cow health in Holsteins. *Journal of Dairy Science* 73: 31-32 [doi: 10.3168/jds.S0022-0302(90)79002-9]
143. Heuer C, Schukken YH and Dobbelaar P (1999). Post-partum body condition score and results from the first test day milk as predictors of disease, fertility, yield and culling in commercial dairy herds. *Journal of Dairy Science* 82: 295- 304 [doi: 10.3168/jds.S0022-0302(99)75236-7]
144. Gillund P, Reksen O, Grohn YT and Karlberg K (2001). Body condition related to ketosis and reproductive performance in Norwegian dairy cows. *Journal of Dairy Science* 84: 1390-1396 [doi: 10.3168/jds.S0022-0302(01)70170-1]
145. Herdt TH (2020). Overview of ketosis in cattle. [msdsvetmanual.com/metabolic-disorders/ketosis-in-cattle/overview-of-ketosis-in-cattle](https://www.msdsvetmanual.com/metabolic-disorders/ketosis-in-cattle/overview-of-ketosis-in-cattle)
146. Ferguson JD, Galligan DT and Thomsen N (1994). Principal descriptors of body condition score in Holstein cows. *Journal of Dairy Science* 77: 2695-2703 [doi: 10.3168/jds.S0022-0302(94)77212-X]
147. Heuer C, Schukken YH and Dobbelaar P (1999). Post-partum body condition score and results from the first test day milk as predictors of disease, fertility, yield and culling in commercial dairy herds. *Journal of Dairy Science* 82: 295-304 [doi: 10.3168/jds.S0022-0302(99)75236-7]

## Sub-clinical milk fever and ketosis in crossbred cows

148. Ruegg PL and Milton RL (1995). Body condition score of Holstein cows on Price Edward Island, Canada: Relationships with yield, reproductive performance and disease. *Journal of Dairy Science* 78: 552-564 [doi: 10.3168/jds.S0022-0302(95)76666-8]
149. Kida K (2003). Relationships of metabolic profiles to milk production and feeding in dairy cows. *Journal of Veterinary Medicine Sciences* 65: 671-677 [doi: 10.1292/jvms.65.671]
150. Kronfeld DS (1971). Hypoglycemia in ketotic cows. *Journal of Dairy Science* 54: 949-961 [doi: 10.3168/jds.S0022-0302(71)85951-9]
151. Jorritsma R, Wensing T, Krup TA, Vos PL and Noordhuizen JP (2003). Metabolic changes in early lactation and impaired reproductive performance in dairy cows. *Veterinary Research* 34: 11-26 [doi: 10.1051/vetres:2002054]
152. Bertics SJ, Grummer RR, Cadorniga-Valino C and Stoddard EE (1992). Effect of prepartum dry matter intake on liver triglyceride concentration and early lactation. *Journal of Dairy Science* 75: 1914-1922 [doi: 10.3168/jds.S0022-0302(92)77951-X]
153. Fiore E, Piccione G, Rizzo M, Morgante M, Barberio A, Giudice E and Ganesella M (2017). Adaptation of some energetic parameters during transition period in dairy cows. *Journal of Applied Animal Research* 46: 402-405 [doi: 10.1080/09712119.2017.1313742]
154. Sakha M, Ameri M and Rohbakhsh A (2006). Changes in blood  $\beta$ -hydroxybutyrate and glucose concentrations during dry and lactation periods in Iranian Holstein cows. *Comparative Clinical Pathology* 15: 221-226 [doi: 10.1007/s00580-006-0650-2]
155. Wathes DC, Cheng Z, Chowdhury W, Fenwick MA, Fitzpatrick R, Morris DG, Patton J and Murphy JJ (2009). Negative energy balance alters global gene expression and immune responses in the uterus of post-partum dairy cows. *Physiology Genomics* 39: 1-13 [doi: 10.1152/physiolgenomics.00064.2009]
156. Holtenius P and Holtenius K (1996). New aspects of ketone bodies in energy metabolism of dairy cows: a review. *Zentralbl Veterinarmed A* 43: 579-587 [doi: 10.1111/j.1439-0442.1996.tb00491.x]
157. Ospina PA, Nydam DV, Stokol T and Overton TR (2010). Associations of elevated nonesterified fatty acids and beta-hydroxybutyrate concentrations with early lactation reproductive performance and milk production in transition dairy cattle in the northeastern United States. *Journal of Dairy Science* 93: 1596-1603 [doi: 10.3168/jds.2009-2852]
158. Kessel S, Stroehl M, Meyer HH, Hiss S, Sauerwein H et al (2008). Individual variability in physiological adaptation to metabolic stress during early lactation in dairy cows kept under equal conditions. *Journal of Animal Science* 86: 2903-2912 [doi: 10.2527/jas.2008-1016]
159. McArt JAA, Nydam DV, Oetzel GR, Overton TR and Ospina PA (2013). Elevated non-esterified fatty acids and  $\beta$ -hydroxybutyrate and their association with transition dairy cow performance. *Veterinary Journal* 198: 560-570 [doi: 10.1016/j.tvjl.2013.08.011]

160. Peterson R and Waldern D (1981). Repeatabilities of serum constituents in Holstein Friesians affected by feeding, age, lactation and pregnancy. *Journal of Dairy Science* 64: 822-831 [doi: 10.3168/jds.S0022-0302(81)82653-7]
161. Hadzimusic N and Krnic J (2012). Values of calcium, phosphorus and magnesium concentrations in blood plasma of cows in dependence on the reproductive cycle and season. *Journal of Faculty of Veterinary Medicine Istanbul University* 38: 1-8
162. Thilising T, Larsen T, Jorgensen RJ and Houe H (2007). The effect of dietary calcium and phosphorus supplementation in zeolite a treated dry cows on peri-parturient calcium and phosphorus homeostasis. *Journal of Veterinary Medicine* 54: 82-91 [doi: 10.1111/j.1439-0442.2007.00887.x]
163. Taylor MS, Knowlton KF, McGilliard ML, Seymour WM and Herbein JH (2008). Blood mineral, hormone and osteocalcin responses of multiparous Jersey cows to an oral dose of 25-hydroxyvitamin D3 or vitamin D3 before parturition. *Journal of Dairy Science* 91: 2408-2416 [doi: 10.3168/jds.2007-0750]
164. Zhang Z, Liu G, Li X, Wang Z, Kong T, Zhang N and Guo C (2009).  $\beta$ -hydroxybutyrate, glucose, calcium, phosphorus and vitamin C concentrations in blood of dairy cows with subclinical ketosis during the early lactation. *Bulletin of the Veterinary Institute in Pulawy* 53: 71-74
165. Sampson JD, Spain JN, Jones C and Carstensen L (2009). Effects of calcium chloride and calcium sulfate in an oral bolus given as a supplement to post-partum dairy cows. *Veterinary Therapeutics* 10: 131-139
166. Blanc CD, Van der List M, Aly SS, Rossow HA and Silva-del-Rio N (2014). Blood calcium dynamics after prophylactic treatment of subclinical hypocalcemia with oral or intravenous calcium. *Journal of Dairy Science* 97: 6901-6906 [doi: 10.3168/jds.2014-7927]
167. Oetzel GR and Goff JP (2008). Milk fever (Parturient paresis) in cows, ewes, and doe goats. Chapter 33. In: Anderson DE and Rings M (2008). *Current Veterinary Therapy- E-Book: Food Animal Practice*. Elsevier Health Sciences, Saunders Elsevier, St. Louis, Missouri 63146, USA pp.130-134
168. Anderson DE and Rings M (2008). *Current Veterinary Therapy- E-Book: Food Animal Practice*. Elsevier Health Sciences, Saunders Elsevier, St. Louis, Missouri 63146, USA
169. Shock DA, Roche SM, Gemore R and Olson ME (2019). A pilot study to evaluate the effect of a novel calcium and vitamin D containing oral bolus on serum calcium levels in Holstein dairy cows following parturition. *Veterinary Medicine: Research and Reports* 10151-10158 [doi: 10.2147/VMRR.S219740]
170. Oetzel GR and Miller BE (2012). Effect of oral calcium bolus supplementation on early-lactation health and milk yield in commercial dairy herds. *Journal of Dairy Science* 95: 7051-7065 [doi: 10.3168/jds.2012-5510]
171. Grummer RR (2008). Nutritional and management strategies for the prevention of fatty liver in dairy cattle. *Veterinary Journal* 176: 10-20 [doi: 10.1016/j.tvjl.2007.12.033]